SITE-SPECIFIC UFP QUALITY ASSURANCE PROJECT PLAN

SUPERIOR BARREL AND DRUM SITE

798 Jacob Harris Road Elk Township, New Jersey 08028

Prepared By:

Removal Support Team 2
Weston Solutions, Inc.
East Division
Edison, New Jersey 08837

DC No.: RST 2-02-F-2782 TDD No.: TO-0029-0151 EPA Contract No.: EP-W-06-072

May 2014

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ATTACHMENTS

ATTACHMENT A: Figure 1: Site Location Map

ATTACHMENT B: EPA/ERT SOPs

SOP No.: 2001 - General Field Sampling Guidelines

SOP No.: 2016 - Sediment Sampling

LIST OF ACRONYMS

ADR Automated Data Review

ANSETS Analytical Services Tracking System AOC Acknowledgment of Completion

ASTM American Society for Testing and Materials

CEO Chief Executive Officer

CERCLA Comprehensive Environmental Response, Compensation and Liability Act

CLP Contract Laboratory Program
CFM Contract Financial Manager

CO Contract Officer
COI Conflict of Interest
COO Chief Operations Officer

CRDL Contract Required Detection Limit
CRTL Core Response Team Leader

CRQL Contract Required Quantitation Limit

CQLOSS Corporate Quality Leadership and Operations Support Services

CWA Clean Water Act

DCN Document Control Number

DESA Division of Environmental Science and Assessment

DI Deionized Water
DPO Deputy Project Officer
DQI Data Quality Indicator
DQO Data Quality Objective
EM Equipment Manager

EDD Electronic Data deliverable ENVL Environmental Unit Leader

EPA Environmental Protection Agency
ERT Environmental Response Team

FASTAC Field and Analytical Services Teaming Advisory Committee

GC/ECD Gas Chromatography/Electron Capture Detector

GC/MS Gas Chromatography/Mass Spectrometry

HASP Health and Safety Plan
HRS Hazard Ranking System
HSO Health and Safety Officer

ITM Information Technology Manager

LEL Lower Explosive Limit
MSA Mine Safety Appliances

MS/MSD Matrix Spike/Matrix Spike Duplicate

NELAC National Environmental Laboratory Accreditation Conference
NELAP National Environmental Laboratory Accreditation Program
NIOSH National Institute for Occupational Safety and Health

NIST National Institute of Standards and Technology

OSC On-Scene Coordinator

OSHA Occupational Safety and Health Administration
OSWER Office of Solid Waste and Emergency Response

LIST OF ACRONYMS (Concluded)

PARCCS Precision, Accuracy, Representativeness, Completeness, Comparability,

Sensitivity

PAH Polynuclear Aromatic Hydrocarbons

PCB Polychlorinated Biphenyls
PIO Public Information Officer

PM Program Manager PO Project Officer

PRP Potentially Responsible Party

PT Proficiency Testing QA Quality Assurance

QAL Quality Assurance Leader
QAPP Quality Assurance Project Plan
QMP Quality Management Plan

QA/QC Quality Assurance/Quality Control

QC Quality Control

RC Readiness Coordinator

RCRA Resource Conservation and Recovery Act

RPD Relative Percent Difference

RSCC Regional Sample Control Coordinator

RST Removal Support Team

SARA Superfund Amendments and Reauthorization Act

SEDD Staged Electronic Data Deliverable

SOP Standard Operating Practice

SOW Statement of Work SPM Site Project Manager

START Superfund Technical Assessment and Response Team

STR Sampling Trip Report
TAL Target Analyte List
TCL Total Compound List

TDD Technical Direction Document
TDL Technical Direction Letter

TO Task Order

TQM Total Quality Management
TSCA Toxic Substances Control Act

UFP Uniform Federal Policy
VOA Volatile Organic Analysis

CROSSWALK

The following table provides a "cross-walk" between the QAPP elements outlined in the Uniform Federal Policy for Quality Assurance Project Plans (UFP-QAPP Manual), the necessary information, and the location of the information within the text document and corresponding QAPP Worksheet. Any QAPP elements and required information that are not applicable to the project are circled.

| QAI | PP Element(s) and Corresponding Section(s) of UFP-QAPP Manual | Required Information | Crosswalk to QAPP Section | Crosswalk to QAPP Worksheet No. |
|-----|---|---|---------------------------|---------------------------------|
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QAPP Worksheet #1: Title and Approval Page

Title: Site-Specific Quality Assurance Project Plan

Site Name/Project Name: Superior Barrel and Drum Site

| Revision Number: 00 | |
|---|---------------------------------------|
| Revision Date: Not Applicable | • |
| Weston Solutions, Inc. | |
| Lead Organization | |
| Patrick Ahern | |
| Weston Solutions, Inc. | |
| 1090 King Georges Post Road, Suite 201 | · |
| Edison, NJ 08837 | |
| Email: Patrick.Ahern@westonsolutions.com | |
| Preparer's Name and Organizational Affiliation | |
| | |
| | |
| May 1, 2014 | |
| Preparation Date (Day/Month/Year) | |
| | 7 |
| Cita Duniant Managan | 111 |
| Site Project Manager: | Jan John |
| Determination of a National Control of the Control | Signature |
| Peter Lisichenko/Weston Solutions, Inc. | |
| Printed Name/Organization/Date | |
| QA Officer/Technical Reviewer: | Anda Co late |
| Q'I Officell Technical Reviewer. | Simolary Sunday |
| Smita Sumbaly/Weston Solutions, Inc. | Signature |
| | |
| Printed Name/Organization/Date | |
| EPA, Region II On-Scene Coordinator (OSC): | |
| (000) | Signature |
| Keith Glenn/EPA, Region II | Signaturo |
| Printed Name/Organization/Date | |
| | , |
| EPA, Region II Quality Assurance Officer (QAO): | |
| | Signature |
| | Signature |
| Printed Name/Organization/Date | |
| - Burning Date | * |
| | · · · · · · · · · · · · · · · · · · · |
| Document Control Number DST 2 02 E 2792 | |

OAPP Worksheet #2: OAPP Identifying Information

Site Name/Project Name: Superior Barrel and Drum Site

Site Location: 798 Jacob Harris Road, Elk Township, Gloucester County, New Jersey

Operable Unit: 00

Title: Site Quality Assurance Project Plan

Revision Number: 00

Revision Date: Not Applicable

- 1. Identify guidance used to prepare QAPP: Uniform Federal Policy for Quality Assurance Project Plans. Refer to EPA/ERT SOPs and DESA Laboratory Methods
- 2. Identify regulatory program: EPA, Region II
- 3. Identify approval entity: EPA, Region II
- 4. Indicate whether the QAPP is a generic or a Site-specific QAPP.
- 5. List dates of scoping sessions that were held: March 28 and April 24, 2014
- 6. List dates and titles of QAPP documents written for previous site work, if applicable:

Site Specific Quality Assurance Project Plan (QAPP), Superior Barrel and Drum, RST 2-02-F-2541, September 6, 2013

- 7. List organizational partners (stakeholders) and connection with lead organization: None
- 8. List data users: EPA, Region II (see Worksheet #4 for individuals)
- 9. If any required QAPP elements and required information are not applicable to the project, then provide an explanation for their exclusion below: None
- 10. Document Control Number: RST 2-02-F-2782

QAPP Worksheet #3: Distribution List

[List those entities to which copies of the approved QAPP, subsequent QAPP revisions, addenda, and amendments are sent]

| QAPP Recipient | Title | Organization | Telephone Number | Fax Number | E-mail Address | Document Control Number |
|------------------|------------|----------------------------------|---------------------|----------------|--------------------------------------|----------------------------|
| Keith Glenn | OSC | EPA, Region II | (732) 452-6415 | (732) 906-6182 | Glenn.keith@epa.gov | RST 2-02-F-2782 |
| Timothy Benton | HSO | Weston Solutions, Inc., RST 2 | (732) 585-4425 | (732) 225-7037 | Tim.Benton@westonsolutions.com | RST 2-02-F-2782 |
| Smita Sumbaly | QA Officer | Weston Solutions, Inc., RST 2 | (732) 585-4410 | (732) 225-7037 | S.Sumbaly@westonsolutions.com | RST 2-02-F-2782 |
| Peter Lisichenko | SPM | Weston Solutions, Inc., RST 2 | (732)-585-4411 | (732) 225-7037 | Peter.Lisichenko@westonsolutions.com | RST 2-02-F-2782 |

QAPP Worksheet #4: Project Personnel Sign-Off Sheet

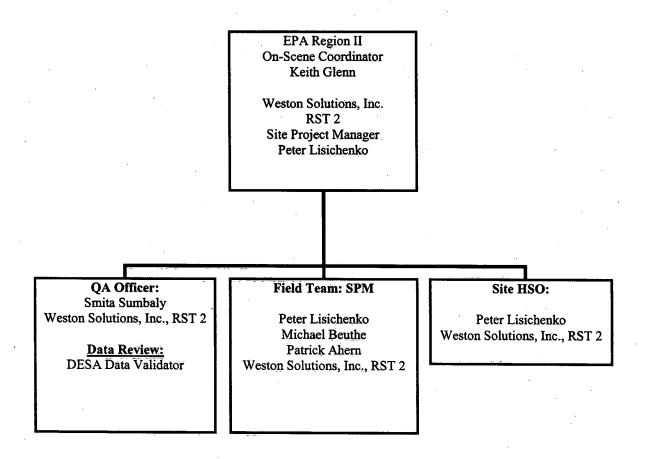
[Copies of this form signed by key project personnel from each organization to indicate that they have read the applicable sections of the QAPP and will perform the tasks as described; add additional sheets as required. Ask each organization to forward signed sheets to the central project file.]

Organization: Weston Solutions, Inc.

| Project Personnel | Title | Telephone Number | Signature | Date QAPP Read |
|-------------------|--|---------------------|---------------|-------------------|
| Keith Glenn | EPA, Region II, On- Scene Coordinator | (732) 321-4454 | | · |
| Peter Lisichenko | Site Project Manager, RST 2 | (732) 585-4411 | Du hum | 5/1/14 |
| Smita Sumbaly | QAO, RST 2 | (732) 585-4410 | Atrita Sumbas | 5/1/14 |
| Timothy Benton | HSO, RST 2 | (732) 585-4425 | 31/2 | 5/1/14 |
| Michael Beuthe | Field Personnel, RST 2 | (732) 585-4447 | | |
| Patrick Ahern | Field Personnel, RST 2 | (732) 585-4448 | 1 | |

QAPP Worksheet #5: Project Organizational Chart

Identify reporting relationship between all organizations involved in the project, including the lead organization and all contractor and subcontractor organizations. Identify the organizations providing field sampling, on-site and off-site analysis, and data review services, including the names and telephone numbers of all project managers, project team members, and/or project contacts for each organization.



Acronyms:

SPM: Site Project Manager HSO: Health & Safety Officer

DESA: Division of Environmental Science and Assessment

QAPP Worksheet #6: Communication Pathways

| Communication Drivers | Responsible Entity | Name | Phone Number | Procedure |
|--------------------------------------|--|--------------------------|----------------|--|
| Point of contact with EPA OSC | Site Project Manager, Weston Solutions, Inc., RST 2 | Peter Lisichenko, SPM | (732) 585-4411 | All technical, QA and decision-making matters in regard to the project (verbal, written or electronic) |
| Adjustments to QAPP | Site Project Manager, Weston Solutions, Inc., RST 2 | Peter Lisichenko, SPM | (732) 585-4411 | QAPP approval dialogue |
| Health and Safety On-Site Meeting | Site Project Manager, Weston Solutions, Inc., RST 2 | Peter Lisichenko, SPM | (732) 585-4411 | Explain Site hazards, personnel protective equipment, hospital location, etc. |

OSC: On-Scene Coordinator SPM: Site Project Manager

QAPP Worksheet #7: Personnel Responsibilities and Qualifications Table

| Name | Title | Organizational Affiliation | Responsibilities | Education and Experience Qualifications* |
|-----------------------|-----------------------------|-------------------------------|---|---|
| Keith Glenn | EPA On-Scene Coordinator | EPA, Region 2 | All project coordination, direction and decision making. | NA |
| Peter Lisichenko, SPM | Site Project Manager, RST 2 | Weston Solutions, Inc. | Implementing and executing the technical, QA and health and safety during sampling event and sample management. | 13 years* |
| Michael Beuthe | Field Personnel, RST 2 | Weston Solutions, Inc. | Sample collection | 3 years* |
| Patrick Ahern | Field Personnel, RST 2 | Weston Solutions, Inc. | Sample collection | 3 years* |

^{*}All RST 2 members, including subcontractor's resumes are in possession of RST 2 Program Manager, EPA Project Officer and Contracting officers.

QAPP Worksheet #8: Special Personnel Training Requirements Table

| Project Function | Specialized Training By Title or Description of Course | Training Provider | Training Date | Personnel / Groups Receiving Training | Personnel Titles / Organizational Affiliation | Location of Training Records / Certificates | | |
|-------------------------------|--|--|---------------------------------------|--|---|--|--|--|
| | [Specify location of training records and certificates for samplers] | | | | | | | |
| QAPP Training | This training is presented to all RST 2 personnel to introduce the provisions, requirements, and responsibilities detailed in the UFP QAPP. The training presents the relationship between the site-specific QA Project Plans (QAPPs), SOPs, work plans, and the Generic QAPP. QAPP refresher training will be presented to all employees following a major QAPP revision. | Weston Solutions, Inc., QAO | As needed | All RST 2 field personnel upon initial employment and as refresher training | Weston Solutions, Inc. | Weston Solutions, Inc., EHS Database | | |
| Health and Safety Training | Health and safety training will be provided to ensure compliance with Occupational Safety and Health Administration (OSHA) as established in 29 CFR 1910.120. | Weston Solutions, Inc., HSO | Yearly at a minimum | All Employees upon initial employment and as refresher training every | Weston Solutions, Inc. | Weston Solutions, Inc., EHS Database | | |
| Others | FORMS II Lite, Scribe, ICS 100 and 200, and Air Monitoring Equipment Trainings provided to all employees | Weston Solutions, Inc., QAO/Group Leader's | Upon initial employment and as needed | year | | | | |
| | Dangerous Goods Shipping | Weston Solutions, Inc., HSO | Every 2 years | | | : | | |

All team members are trained in the concepts and procedures in recognizing opportunities for continual improvement, and the approaches required to improve procedures while maintaining conformance with legal, technical, and contractual obligations.

¹All RST 2 members, including subcontractor's certifications are in possession of RST 2 HSO.

QAPP Worksheet #9: Project Scoping Session Participants Sheet

Site Name/Project Name: Superior Barrel and Drum Site

Site Location: 798 Jacob Harris Road, Elk Township, Gloucester County, New Jersey

Operable Unit: 00

Date of Session: March 28 and April 24, 2014

Scoping Session Purpose: To discuss questions, comments, and assumptions regarding

technical issues involved with the sampling activities.

| Name | Title | Affiliation | Phone # | E-mail Address | *Project Role |
|------------------|-------------------------|----------------------------------|----------------|---------------------------------------|----------------------------|
| Keith Glenn | On-Scene Coordinator | EPA, Region II | (732) 452-6415 | Glenn.keith@epa.gov | OSC |
| Peter Lisichenko | Site Project Manager | Weston Solutions, Inc., RST 2 | (732) 585-4411 | Peter.Lisichenko@westons olutions.com | Site Project Management |

Comments/Decisions:

As part of the Removal Assessment, Weston Solutions, Inc., Removal Support Team 2 (RST 2) is tasked by the EPA to sample sediments located at both on-site and off-site locations, including wetland areas. Twenty Sediment samples will be collected via plastic scoops and En Core volumetric sampling devices. Samples will be submitted to the EPA DESA laboratory for (TCL) volatile organic compounds (VOC), semi-volatile organic compound (SVOC), polychlorinated biphenyl (PCB), pesticides, target analyte list (TAL) metal, mercury, and cyanide analyses. The samples will be collected for definitive data QA objectives. One field duplicate sample and additional volume for MS/MSD samples will be collected. One trip blank sample will be included in the sample cooler for the first set of samples collected.

Action Items:

RST 2 Submitted a CLP request form for analytical services on April 24, 2014. Analytical services request approval was received on April 28, 2014 for EPA DESA laboratory.

Consensus Decisions:

Sampling conducted as part of the Removal Assessment will begin on May 5, 2014 and last approximately for two days

QAPP Worksheet #10: Problem Definition

PROBLEM DEFINITION

Based up on New Jersey Department of Environmental Protection's (NJDEP) observations from August 2013 and subsequent observations by EPA; thousands of containers, mostly 275-gallon totes and 55-gallon drums, located along the road as well as in the woods, wetlands, and elsewhere throughout the Site. Containers were shown to be various states of deterioration. They were found to be leaking, void of tops, exposed to weather elements, rusted, damaged due to gunshots, stored improperly, and laying on their sides. Numerous trailers were also found to be open and containing 55-gallon drums. The containers throughout the Site appeared to be full of contents, however most did not have labels. Labels on some containers include flammable liquids, corrosive, marine pollutant, flammable solid and non-hazardous material. Based on NJDEP's field screening, the materials did not reflect the labels on the containers for many of drums. Solid and Liquid Waste sampling was conducted by Weston Solutions, Inc., RST 2 as part of the 2013 Removal Assessment of the Site. The analytical data was used to determine the presence and nature of the contaminants at the Site. As a follow up to the 2013 sampling event, sediment sampling will be conducted by Weston Solutions, Inc., RST 2 at wetland areas located both on-site and the adjoining property. Sampling is scheduled to commence on May 5, 2014 and is anticipated to be completed in two days.

SITE HISTORY/CONDITIONS

The Site is located at 798 Jacob Harris Road in Elk Township, Gloucester County, New Jersey (coordinates 39.6869, -75.132314). The facility consists of a main processing building and numerous trailers located throughout the 5.5 acre property. North of the Site is bordered by Industrial Drum Company and a chain-link fence separates the two properties. Jacob Harris Road marks the eastern boundary of the Site, beyond which is a densely forested property. To the South are private lands which are also densely wooded with several marshy areas. The Western boundary is indicated by Rt. 55, a major highway. Currently, the facility is inoperable with last known operation activity occurring in 2012. The Site is unsecured from each direction and evidence of trespassers has been noted. All doors of the main building and trailers are open.

The Site consists of two operational areas. The main area is where the permanent steel structure is located. This area would receive containers, rinse the containers, and recondition them for future market. This area is approximately 2.4 acres with containers located throughout. The additional operational area appears to be mainly storage of full 275-gallon and 55-gallon totes, with several trailers holding containers. This area encompasses approximately 0.32 acres of land, also holding hundreds of totes and drums. Both areas show signs of impact from leaking containers or dumping of materials.

EPA conducted a Removal Assessment at the Site in September 2013. Waste samples were collected from the drums, tanks, and other containers on site, as well as surface water and soil samples. Samples were screened in the field by EPA's Emergency and Rapid Response Services (ERRS) contractor using a HazCat field screening kit. Field screening results indicated that many of these contained housed hazardous materials, including flammables and corrosives. More than 100 of these samples were sent to a laboratory for target compound list (TCL) VOC, semi-

QAPP Worksheet #10: Problem Definition (Concluded)

volatile organic compound (SVOC), polychlorinated biphenyl (PCB), pesticide, target analyte list (TAL) metal, mercury, and cyanide analyses. Resulting from this sampling indicated high levels of VOCs and SVOCs within many of the containers, as well as pesticides and PCBs in some of the containers. Results also indicated that contamination of some of the on-site soils had occurred.

PROJECT DESCRIPTION

In order to assist EPA with the assessment activities, RST 2 is tasked with the collection of 20 sediment samples from on-site and adjoining wetland areas. The sediment sampling will start on May 5, 2014 and conclude on May 6, 2014. Samples will be collected along four transects located both on and off the subject site with approximately five samples taken from each. Samples will be submitted to the EPA DESA laboratory for (TCL) volatile organic compounds (VOC), semi-volatile organic compound (SVOC), polychlorinated biphenyl (PCB), pesticides, target analyte list (TAL) metal, mercury, and cyanide analyses. The samples will be collected for definitive data QA objectives. One field duplicate sample and additional volume for MS/MSD samples will be collected. One trip blank sample will be included in the sample cooler for the first set of samples collected.

PROJECT DECISION STATEMENTS

The analytical data from this investigation will be used to assist the EPA in determining the presence and nature of the contaminants at the Site and the adjoining Freshwater Forested/Shrub Wetlands.

QAPP Worksheet # 11: Project Quality Objectives/Systematic Planning Process Statement

Overall project objectives include: RST 2 is tasked with the collection of 20 sediment samples from on-site and adjoining wetland areas. The sediment sampling will start on May 5, 2014 and conclude on May 6, 2014. Samples will be collected along four transects located both on and off the subject site with approximately five samples taken from each. Samples will be submitted to the EPA DESA laboratory for (TCL) volatile organic compound (VOC), semi-volatile organic compound (SVOC), polychlorinated biphenyl (PCB), pesticides, target analyte list (TAL) metal, mercury, and cyanide analyses. The samples will be collected for definitive data QA objectives. One field duplicate sample and additional volume for MS/MSD samples will be collected. One trip blank sample will be included in the sample cooler for the first set of samples collected. The analytical data from this investigation will be used to assist the EPA in determining the presence and nature of the contaminants at the Site and the adjoining Freshwater Forested/Shrub Wetlands.

Who will use the data? Data and field screening characterization will be used by the EPA, Region II OSC.

What will the data be used for? Data from this sampling event will be used to determine the presence of the contaminants in wetland areas located on and off-site

What types of data are needed?

Matrix: Sediment

Type of Data: Definitive data

Analytical Techniques: Off-site laboratory analyses

Parameters: Full TCL (VOCs, SVOCs, pesticides, and PCBs); and TAL metals, including

mercury and cyanide

Type of sampling equipment: Sample jars, En Core Samplers, plastic scoops, aluminum pie

tins, and zip-lock bags

Access Agreement: Obtained by EPA, Region II OSC

Sampling locations: Sampling will be done along three parallel transects that start near the central portion of the site and extend south/southeast beyond the property boundary and into the adjoining wetland areas. A fourth separate sampling transect line will be located offsite, southeast of the subject property.

How much data are needed? Twenty sediment samples will be collected from the sampling transects located both on and offsite. Approximately 5 samples will be collected from each transect line. Samples will be submitted to the EPA DESA laboratory for (TCL) volatile organic compound (VOC), semi-volatile organic compound (SVOC), polychlorinated biphenyl (PCB), pesticides, target analyte list (TAL) metal, mercury, and cyanide analyses. The samples will be collected for definitive data QA objectives. One field duplicate sample and additional volume for MS/MSD samples will be collected. One trip blank sample will be included in the sample cooler for the first set of samples collected.

How "good" does the data need to be in order to support the environmental decision?

Sampling/analytical measurement performance criteria for Precision, Accuracy, Representativeness, Completeness, and Comparability (PARCC) parameters will be established. Refer to Worksheet #12, criteria for performance measurement for definitive data.

QAPP Worksheet # 11: Project Quality Objectives/Systematic Planning Process Statement (Concluded)

Where, when, and how should the data be collected/generated? Sample locations will be selected by EPA OSC along the designated transect lines to determine overall condition of the wetland areas located on Site and on adjoining property. All samples will be collected utilizing methods outlined in the Standard Operating Procedures (SOPs). GPS data for all sample locations will be collected. The Removal Assessment is scheduled to begin on May 5, 2014 and will last approximately two days.

Who will collect and generate the data? The sediment samples will be collected by RST 2 and analyzed by EPA DESA laboratory for VOC, SVOC, TAL-metals (including mercury and cyanide), pesticides and PCB analyses. The results obtained will be validated by EPA DESA data validators.

How will the data be reported? All data will be reported by the assigned laboratories (Preliminary, Electronics, and Hard Copy format). The Site Project Manager will provide a Sampling Trip Report, Status Reports, Maps/Figures, Analytical Report, and Data Validation Report to the EPA OSC.

How will the data be archived? Electronic data deliverables will be archived in the Scribe database.

QAPP Worksheet #12: Measurement Performance Criteria Table Worksheet # 12A: Volatile - Organics (DESA)

| Matrix | Sediment/Aqueous ¹ | 1 | | | |
|---------------------|---------------------------------------|-----------------------------------|--|--|---|
| Analytical Group | VOA | | • | | |
| Concentration Level | Medium | | | | |
| Sampling Procedure | Analytical Method/SOP ³ | Data Quality Indicators (DQIs) | Measurement Performance Criteria | QC Sample and/or Activity Used to Assess Measurement Performance | QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A) |
| SOP 2016 | See worksheets:#28/ #23 | Precision Accuracy | % RPD < 20 Average Recovery 70- 130% | LCS Duplicate | A |
| | | Accuracy | Factor of two(-50% to + 100%) from the initial/continuing calibration | Internal standards | Α |
| | | Accuracy | Compound Specific (full range: 17-259%) | Matrix spike | Α |
| | | Accuracy | Limits 70%- 130%(Aqueous) Table 7 of C-123(low Soil) | Surrogate Compounds | Α |
| | | Accuracy | < RL | Method Blank | A |

SOP – Sample Operating Procedure

Aqueous sample will consist of a trip blank sample only. Aqueous field duplicate and MS/MSD will not be collected

QAPP Worksheet #12: Measurement Performance Criteria Table (Continued) Worksheet # 12B: Semivolatile - Organics (DESA)

| Matrix | Sediment | | | | |
|---------------------------------|---------------------------------------|--------------------------------|---|--|---|
| Analytical Group ¹ | Semi-Volatiles | | | | · |
| Concentration Level | Medium | | | | |
| Sampling Procedure ² | Analytical Method/SOP ³ | Data Quality Indicators (DQIs) | Measurement Performance Criteria | QC Sample and/or Activity Used to Assess Measurement Performance | QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A) |
| SOP 2016 | See worksheet #28 & #23 | Precision | % RPD < 30 | LCS Duplicate | A |
| | | Accuracy | Compound Specific (full range: D-262%) | | |
| | | Accuracy | Factor of two(-50% to + 100%) from the initial/continuing calibration | Internal standards | A |
| | | Accuracy | Compound Specific (full range: D-262%) | Matrix spike | A |
| | | Accuracy | Limits 30%-120% for Base Neutrals Limits 20%-120% for Acids | Surrogate Compounds | A |
| | | Accuracy | < RL | Method Blank | , A |

SOP - Sample Operating Procedure

QAPP Worksheet #12 Measurement Performance Criteria Table (Continued) Worksheet # 12C: Pesticides/PCB – Organics (DESA)

| Matrix | Sediment | | | | • |
|---------------------------------|---------------------------------------|--------------------------------|--|--|---|
| Analytical Group ¹ | Pest/PCB | , | | · · · · · · · · · · · · · · · · · · · | |
| Concentration Level | Medium | | | | |
| Sampling Procedure ² | Analytical Method/SOP ³ | Data Quality Indicators (DQIs) | Measurement Performance Criteria | QC Sample and/or Activity Used to Assess Measurement Performance | QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A) |
| SOP 2016 | See worksheets #28/ #23 | Precision Accuracy | % RPD < 30 Average Recovery 50- 150% | LCS Duplicate | A |
| | | Accuracy | Compound Specific (full range: 30-150%) | Matrix spike | A |
| | | Accuracy | Limits 30%-150% | Surrogate Compounds | A |
| | | Accuracy | < RL | Method Blank | Α |
| | | | | | |

SOP - Sample Operating Procedure

QAPP Worksheet #12 Measurement Performance Criteria Table (Continued) Worksheet # 12D: TAL Metals - Inorganics (DESA)

| Matrix | Sediment | · | | | |
|---------------------------------|---------------------------------------|-----------------------------------|--|--|---|
| Analytical Group | Metals/Mercury/Cyanide | | | | |
| Concentration Level | Medium | | | | |
| Sampling Procedure ² | Analytical Method/SOP ³ | Data Quality Indicators (DQIs) | Measurement Performance Criteria | QC Sample and/or Activity Used to Assess Measurement Performance | QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A) |
| SOP 2016 | See #28/ #23 | Precision | % RPD < 20(Aq), % RPD <25(Soil) | LCS Duplicate | A |
| | | Accuracy | Limits: Average Recovery ± 20% aqueous, ± 25% Soil) | LCS | A |
| | | Accuracy | ± 20% aqueous, ± 25% Soil) | Matrix spike | Α |
| | | Precision | < RL Except for Al, Fe, Ca, K, Mg and Na | Interference Check Sample(ICP/AES) | A |
| | | Accuracy | < RL | Method Blank | A |
| | | Precision | RPD < 20 % | Serial Dilution Test(ICP/AES) | A |
| | · | Accuracy | Range of 0.60-1.87 of the original response in the calibration blank | Internal Standards(ICP-MS) | A |

SOP - Sample Operating Procedure

QAPP Worksheet #13: Secondary Data Criteria and Limitations Table

Any data needed for project implementation or decision making that are obtained from non-direct measurement sources such as computer databases, background information, technologies and methods, environmental indicator data, publications, photographs, topographical maps, literature files and historical data bases will be compared to the DQOs for the project to determine the acceptability of the data. Thus, for example, analytical data from historical surveys will be evaluated to determine whether they satisfy the validation criteria for the project and to determine whether sufficient data was provided to allow an appropriate validation to be done. If not, then a decision to conduct additional sampling for the site may be necessary.

| Secondary Data | Data Source (Originating Organization, Report Title, and Date) | Data Generator(s) (Originating Org., Data Types, Data Generation/ Collection Dates) | How Data May Be Used (if deemed usable during data assessment stage) | Limitations on Data Use |
|---------------------------|---|---|--|-------------------------|
| UFP HASP, October 2013 | RST 2 Site - Specific HASP, Superior Barrel and Drum Final Trip Report | Weston Solutions, Inc. | Data used to confirm soil contamination. | N/A |

QAPP Worksheet #14: Summary of Project Tasks

Sampling Tasks:

As part of the Removal Assessment, Weston Solutions, Inc., Removal Support Team 2 (RST 2) is tasked with the collection of 20 sediment samples from on-site and adjoining wetland areas. The sediment sampling will start on May 5, 2014 and conclude on May 6, 2014. Samples will be collected along four transects located both on and off the subject site with approximately five samples taken from each. Samples will be submitted to the EPA DESA laboratory for (TCL) volatile organic compound (VOC), semi-volatile organic compound (SVOC), polychlorinated biphenyl (PCB), pesticides, target analyte list (TAL) metal, mercury, and cyanide analyses. The samples will be collected for definitive data QA objectives. One field duplicate sample and additional volume for MS/MSD samples will be collected. One trip blank sample will be included in the sample cooler for the first set of samples collected. The analytical data from this investigation will be used to assist the EPA in determining the presence and nature of the contaminants at the Site and the adjoining Freshwater Forested/Shrub Wetlands.

Analysis Tasks:

Sediment – TCL VOCs – DESA Method C-123

Sediment – TCL SVOCs – DESA Method C-90

Sediment – TCL PCBs – DESA Method C-91

Sediment - TCL Pesticides - DESA Method C-91

Sediment - TAL Metals (including mercury and Cyanide) - DESA Method C-109, C-110, and C-28

Quality Control Tasks:

All samples will be collected for Definitive Data QA Objective. QA/QC samples will include the collection of one field duplicate for sediment samples, and any additional volume for MS/MSD at the ratio of 1 per 20 samples and 1 per matrix.

Data Management Tasks:

Activities under this project will be reported in status and trip reports and other deliverables (e.g., analytical reports, final reports) described herein. Activities will also be summarized in appropriate format for inclusion in monthly and annual reports.

The following deliverables will be provided under this project:

Maps/Figures: Maps depicting site layout and sample locations will be included in the final report.

QAPP Worksheet #14: Summary of Project Tasks (Continued)

Analytical Report: An analytical report will be prepared for samples analyzed under this plan. Information regarding the analytical methods or procedures employed, sample results, QA/QC results, chain-of-custody documentation, laboratory correspondence, and raw data will be provided within this deliverable.

<u>Data Review:</u> A review of the data generated under this plan will be undertaken. The assessment of data acceptability or usability will be provided separately, or as part of the analytical report.

<u>Final Report</u>: A final report will be prepared which will include a description of field activities, analysis of laboratory results and field data, a sample location map, photo log, and data tables.

Documentation and Records:

All sample documents will be completed legibly, in ink. Any corrections or revisions will be made by lining through the incorrect entry and by initialing the error.

<u>Field Logbook:</u> The field logbook is essentially a descriptive notebook detailing site activities and observations so that an accurate account of field procedures can be reconstructed in the writer's absence. Field logbook will be bound and paginated. All entries will be dated and signed by the individuals making the entries, and should include (at a minimum) the following:

- 1. Site name and project number
- 2. Name(s) of personnel on-site
- 3. Dates and times of all entries (military time preferred)
- 4. Descriptions of all site activities, site entry and exit times
- 5. Noteworthy events and discussions
- 6. Weather conditions
- 7. Site observations
- 8. Sample and sample location identification and description*
- 9. Subcontractor information and names of on-site personnel
- 10. Date and time of sample collections, along with chain of custody information
- 11. Record of photographs
- 12. Site sketches

<u>Sample Labels</u>: Sample labels will clearly identify the particular sample, and should include the following:

- 1. Site/project number.
- 2. Sample identification number.
- 3. Sample collection date and time.
- 4. Designation of sample (grab or composite).
- 5. Sample preservation.
- 6. Analytical parameters.
- 7. Name of sampler.

^{*} The description of the sample location will be noted in such a manner as to allow the reader to reproduce the location in the field at a later date.

OAPP Worksheet #14: Summary of Project Tasks (Concluded)

Sample labels will be written in indelible ink and securely affixed to the sample container. Tieon labels can be used if properly secured.

<u>Custody Seals:</u> Custody seals demonstrate that a sample container has not been tampered with or opened. The individual in possession of the sample(s) will sign and date the seal, affixing it in such a manner that the container cannot be opened without breaking the seal. The name of this individual, along with a description of the sample packaging, will be noted in the field logbook.

Assessment/Audit Tasks:

No performance audit of field operations is anticipated at this time. If conducted, performance and system audit will be in accordance with the project plan.

Data Review Tasks:

All data will be validated by the DESA Laboratory Data Validators.

Laboratory analytical results will be assessed by the data reviewer for compliance with required precision, accuracy, completeness, representativeness, and sensitivity.

QAPP Worksheet #15A Reference Limits and Evaluation Table

Matrix:

Aqueous (Trip Blank)

Analytical Group:

Target Compound List Volatile Organic Compounds

Concentration Level:

Medium

| | CAS | Project (PRP) Quantitation | | Achievable Laboratory (DESA) Limits | | |
|---|------------|----------------------------------|------------|-------------------------------------|-----------|--|
| Analyte | Number | Limit | Method QLs | MDLs μg/kg | RLs µg/kg | |
| Dichlorodifluoromethane | 75-71-8 | NS | 5 μg/L | 0.3 | 5 μg/L | |
| Chloromethane | 74-87-3 | . NS | 5 μg/L | 0.54 | 5 μg/L | |
| Vinyl Chloride | 75-01-4 | NS | 5 μg/L | 1.52 | 5 μg/L | |
| Bromomethane | 74-83-9 | NS | 5 μg/L | 1.90 | 5 μg/L | |
| Chloroethane | 75-00-3 | NS | 5 μg/L | 1.01 | 5 μg/L | |
| Trichlorofluoromethane | 75-69-4 | NS | 5 μg/L | 2.18 | 5 μg/L | |
| 1,1-Dichloroethene | 75-35-4 | NS | 5 μg/L | 1.12 | 5 μg/L | |
| 1,1,2-Trichloro-1,2,2- trifluoroethane | 76-13-1 | NS | 5 μg/L | 0.3 | 5 μg/L | |
| Carbon Disulfide | 75-15-0 | NS | 5 μg/L | 1.58 | 5 μg/L | |
| Acetone | 67-64-1 | NS | 10 μg/L | 0.67 | 10 μg/L | |
| Methyl Acetate | 79-20-9 | NS | 5 μg/L | 0.4 | 5 μg/L | |
| Methylene Chloride | 75-09-2 | NS | 5 μg/L | 0.52 | 5 μg/L | |
| trans-1,2-Dichloroethene | 156-60-5 | NS | 5 μg/L | 0.91 | 5 μg/L | |
| cis-1,2-Dichloroethene | 156-59-2 | NS | 5 μg/L | 0.2 | 5 μg/L | |
| Methyl tert-Butyl Ether | 1634-04-4 | NS | 5 μg/L | 0.4 | 5 μg/L | |
| 1,1-Dichloroethane | 75-34-3 | NS | 5 μg/L | 0.58 | 5 μg/L | |
| 2-Butanone | 78-93-3 | NS | 10 μg/L | 0.7 | 10 μg/L | |
| Chloroform | 67-66-3 | NS | 5 μg/L | 0.44 | 5 μg/L | |
| 1,2-Dichloroethane | 107-06-2 | NS | 5 μg/L | 0.55 | 5 μg/L | |
| 1,1,1-Trichloroethane | 71-55-6 | NS | 5 μg/L | 0.6 | 5 μg/L | |
| Cyclohexane | 110-82-7 | NS | 5 μg/L | 0.6 | 5 μg/L | |
| Carbon Tetrachloride | 56-23-5 | NS | 5 μg/L | 1.23 | 5 μg/L | |
| Benzene | 71-43-2 | NS | 5 μg/L | 0.46 | 5 μg/L | |
| Trichloroethene | 79-01-6 | NS | 5 μg/L | 0.99 | 5 μg/L | |
| Methylcyclohexane | 108-87-2 | NS | 5 μg/L | 0.7 | 5 μg/L | |
| 1,2-Dichloropropane | 78-87-5 | NS | 5 μg/L | 0.44 | 5 μg/L | |
| Bromodichloromethane | 75-27-4 | NS | 5 μg/L | 0.51 | 5 μg/L | |
| cis-1,3-Dichloropropene | 10061-01-5 | NS | 5 μg/L | 0.63 | 5 μg/L | |
| trans-1,3-Dichloropropene | 10061-02-6 | NS | 5 μg/L | 0.4 | 5 μg/L | |
| 1,1,2-Trichloroethane | 79-00-5 | NS | 5 μg/L | 0.3 | 5 μg/L | |
| Dibromochloromethane | 124-48-1 | NS | 5 μg/L | 0.2 | 5 μg/L | |
| 4-Methyl-2-Pentanone | 108-10-1 | NS | 10 μg/L | 0.64 | 10 μg/L | |
| Toluene | 108-88-3 | NS | 5 μg/L | 0.77 | 5 μg/L | |
| 1,2-Dibromoethane | 106-93-4 | NS | 5 μg/L | 0.2 | 5 μg/L | |

QAPP Worksheet #15A Reference Limits and Evaluation Table (Concluded)

Matrix:

Aqueous (Trip Blank)

Analytical Group:

Target Compound List Volatile Organic Compounds - Concluded

Concentration Level:

Medium

| | CAS | Project (PRP) Quantitation | Method CRQL | Achievable Labo | • • • |
|-----------------------------|-------------|----------------------------------|----------------|-----------------|-----------|
| Analyte | Number | Limit | μg/kg | MDLs μg/kg | RLs μg/kg |
| Chlorobenzene | 108-90-7 | NS | 5 μg/L | 0.59 | 5 μg/L |
| Tetrachloroethene | 127-18-4 | NS | 5 μg/L | 1.11 | 5 μg/L |
| 2-Hexanone | 591-78-6 | NS | 10 μg/L | 0.68 | 10 μg/L |
| Ethylbenzene | 100-41-4 | NS | 5 μg/L | 0.59 | 5 μg/L |
| m,p-Xylene | 179601-23-1 | NS | .5 μg/L | 1.17 | 5 μg/L |
| o-Xylene | 95-47-6 | NS | 5 μg/L | 0.56 | 5 μg/L |
| Styrene | 100-42-5 | NS | 5 μg/L | 0.57 | 5 μg/L |
| Bromoform | 75-25-2 | NS | 5 μg/L | 0.43 | 5 μg/L |
| Isopropylbenzene | 98-82-8 | NS | 5 μg/L | 0.3 | 5 μg/L |
| 1,1,2,2-Tetrachloroethane | 79-34-5 | NS | 5 μg/L | 0.64 | 5 μg/L |
| 1,3-Dichlorobenzene | 541-73-1 | NS | 5 μg/L | 0.82 | 5 μg/L |
| 1,4-Dichlorobenzene | 106-46-7 | NS | 5 μg/L | 0.84 | 5 μg/L |
| 1,2-Dichlorobenzene | 95-50-1 | NS | 5 μg/L | 0.75 | 5 μg/L |
| 1,2-Dibromo-3-Chloropropane | 96-12-8 | NS | 5 μg/L | 0.7 | 5 μg/L |
| 1,2,4-Trichlorobenzene | 120-82-1 | NS | 5 μg/L | 0.4 | 5 μg/L |
| *1,2,3-Trichlorobenzene | 87-61-6 | ŅS | 5 μg/L | | 5 μg/L |
| * Bromochloromethane | 74-97-5 | NS | 5 μg/L | | 5 μg/L |

NS - Not Specified

^{*} MDL study will be performed.

QAPP Worksheet #15B Reference Limits and Evaluation Table

Matrix:

Sediment

Analytical Group:

Volatile Organic Compounds

Concentration Level:

Medium

| | CAS | Project (PRP) Quantitation | Method CRQL | Achievable (DESA) | Limits |
|---|------------|----------------------------------|----------------|-------------------|-----------|
| Analyte | Number | Limit | μg/kg | MDLs μg/kg | RLs μg/kg |
| Dichlorodifluoromethane | 75-71-8 | NS | 250 | NA. | 250 |
| Chloromethane | 74-87-3 | NS | 250 | NA | 250 |
| Vinyl Chloride | 75-01-4 | NS | 250 | NA | 250 |
| Bromomethane | 74-83-9 | NS | 250 | NA NA | 250 |
| Chloroethane | 75-00-3 | NS | 250 | NA | 250 |
| Trichlorofluoromethane | 75-69-4 | NS | 250 | NA | 250 |
| 1,1-Dichloroethene | 75-35-4 | NS | 250 | NA | 250 |
| 1,1,2-Trichloro-1,2,2- trifluoroethane | 76-13-1 | NS | 250 | NA | 250 |
| Carbon Disulfide | 75-15-0 | NS | 250 | NA | 250 |
| Acetone | 67-64-1 | NS | 500 | NÄ | 500 |
| Methyl Acetate | 79-20-9 | NS | 250 | NA | 250 |
| Methylene Chloride | 75-09-2 | NS | 250 | NA | 250 |
| trans-1,2-Dichloroethene | 156-60-5 | NS | 250 | NA | 250 |
| cis-1,2-Dichloroethene | 156-59-2 | NS | 250 | NA | 250 |
| Methyl tert-Butyl Ether | 1634-04-4 | NS | 250 | ŇA | 250 |
| 1,1-Dichloroethane | 75-34-3 | NS | 250 | NA | 250 |
| 2-Butanone | 78-93-3 | NS | 500 | NA | 500 |
| Chloroform | 67-66-3 | NS | 250 | NA | 250 |
| 1,2-Dichloroethane | 107-06-2 | NS | 250 | ŇĀ | 250 |
| 1,1,1-Trichloroethane | 71-55-6 | NS | 250 | NA | 250 |
| Cyclohexane | 110-82-7 | NS | 250 | NA | 250 |
| Carbon Tetrachloride | 56-23-5 | NS | 250 | NA | 250 |
| Benzene | 71-43-2 | - NS | 250 | ŇA | 250 |
| Trichloroethene | 79-01-6 | NS | 250 | NA | 250 |
| Methylcyclohexane | 108-87-2 | NS | 250 | NA | 250 |
| 1,2-Dichloropropane | 78-87-5 | NS | 250 | NA | 250 |
| Bromodichloromethane | 75-27-4 | NS | 250 | NA | 250 |
| cis-1,3-Dichloropropene | 10061-01-5 | NS | 250 | NA | 250 |
| trans-1,3-Dichloropropene | 10061-02-6 | NS | 250 | NA | 250 |
| 1,1,2-Trichloroethane | 79-00-5 | NS | 250 | NA | 250 |
| Dibromochloromethane | 124-48-1 | NS | 250 | NA | 250 |
| 4-Methyl-2-Pentanone | 108-10-1 | NS | 500 | NA | 500 |
| Toluene | 108-88-3 | NS | 250 | NA | 250 |

NS - Not Specified

NA - Not Applicable

QAPP Worksheet #15B Reference Limits and Evaluation Table (Concluded)

Matrix:

Sediment

Analytical Group:

Volatile Organic Compounds

Concentration Level:

Medium

| Analyte | CAS Number | Project (PRP) Quantitation Limit | Method CRQL μg/kg | | Laboratory) Limits RLs µg/kg |
|-----------------------------|---------------|----------------------------------|-------------------------|------|-------------------------------------|
| 1,2-Dibromoethane | 106-93-4 | NS | 250 | NA I | 250 |
| Chlorobenzene | 108-90-7 | NS | 250 | NA | 250 |
| Tetrachloroethene | 127-18-4 | NS | 250 | NÄ | 250 |
| 2-Hexanone | 591-78-6 | NS | 500 | NA | 500 |
| Ethylbenzene | 100-41-4 | NS | 250 | NA | 250 |
| m,p-Xylene | 179601-23-1 | NS | 250 | NA | 250 |
| o-Xylene | 95-47-6 | NS | 250 | NÄ | 250 |
| Styrene | 100-42-5 | NS | 250 | NA | 250 |
| Bromoform | 75-25-2 | NS | 250 | NA | 250 |
| Isopropylbenzene | 98-82-8 | NS | 250 | NA | 250 |
| 1,1,2,2-Tetrachloroethane | 79-34-5 | NS | 250 | NA | 250 |
| 1,3-Dichlorobenzene | 541-73-1 | NS | 250 | NA | 250 |
| 1,4-Dichlorobenzene | 106-46-7 | NS | 250 | NA | 250 |
| 1,2-Dichlorobenzene | 95-50-1 | NS | 250 | NA | 250 |
| 1,2-Dibromo-3-Chloropropane | 96-12-8 | NS | 250 | NA | 250 |
| 1,2,4-Trichlorobenzene | 120-82-1 | NS | 250 | NA | 250 |
| 1,2,3-Trichlorobenzene | 87-61-6 | NS | 250 | NA | 250 |
| Bromochloromethane | 74-97-5 | NS | 250 | NA | 250 |

Note: Based on the new CRQLs the MDL study is currently being reanalyzed.

NS - Not Specified

NA - Not Applicable

QAPP Worksheet #15C Reference Limits and Evaluation Table

Matrix:

Sediment

Analytical Group:

Semi -Volatile Organic Compounds

Concentration Level:

Low/Medium

| | CAS | Project (PRP) Quantitation | Method QLs | Achievable Laboratory (DESA) Limits | |
|-----------------------------|----------|----------------------------|---------------|--|-----------|
| Analyte | Number | Limit ³ | μg/kg | MDLs μg/kg | RLs μg/kg |
| Benzaldehyde | 100-52-7 | NS | 170 | NA | 120 |
| Phenol | 108-95-2 | NS | 170 | NA | 120 |
| Bis(2-Chloroethyl)Ether | 111-44-4 | NS | 170 | NA | 120 |
| 2-Chlorophenol | 95-57-8 | NS | 170 | NA | 120 |
| 2-Methylphenol | 95-48-7 | NS | 170 | NA | 120 |
| Bis(2-Chloroisopropyl)Ether | 108-60-1 | NS | 170 | NA | 120 |
| Acetophenone | 98-86-2 | NS | 170 | NA | 120 |
| 4-Methylphenol | 106-44-5 | NS | 170 | NA | 120 |
| N-Nitroso-Di-N-Propylamine | 621-64-7 | NS | 170 | NA | 120 |
| Hexachloroethane | 67-72-1 | NS | 170 | NA | 120 |
| Nitrobenzene | 98-95-3 | NS | 170 | NA | 120 |
| Isophorone | 78-59-1 | NS | 170 | NA | 120 |
| 2-Nitrophenol | 88-75-5 | NS | 170 | NA NA | 120 |
| 2,4-Dimethylphenol | 105-67-9 | NŠ | 170 | NA | 120 |
| Bis(2-Chloroethoxy)Methane | 111-91-1 | NS | 170 | NA | 120 |
| 2,4-Dichlorophenol | 120-83-2 | NS | 170 | NA | 120 |
| Naphthalene | 91-20-3 | NS | 170 | NA | 120 |
| 4-Chloroaniline | 106-47-8 | ŃŚ | 170 | NA | 120 |
| Hexachlorobutadiene | 87-68-3 | NS | 170 | NA | 120 |
| Caprolactam . | 105-60-2 | NS | 170 | NA | 120 |
| 4-Chloro-3-Methylphenol | 59-50-7 | NS | 170 | NA | 120 |
| 2-Methyl Naphthalene | 91-57-6 | NS | 170 | NA | 120 |
| Hexachlorocyclopentadiene | 77-47-4 | NS | 170 | NA | 120 |
| 1,2,4,5-Tetrachlorobenzene | 95-94-3 | NS | 170 | NA | 120 |
| 2,4,6-Trichlorophenol | 88-06-2 | NS | 170 | NA | 120 |
| 2,4,5-Trichlorphenol | 95-95-4 | NS | 170 | NA | 120 |
| 1,1'-Biphenyl | 92-52-4 | NS | 170 | NA | 120 |
| 2-Chloronaphthalene | 91-58-7 | NS | 170 | NA | 120 |
| 2-Nitroaniline | 88-74-4 | NS | 330 | NA | 120 |
| Dimethyl Phthalate | 131-11-3 | NS | 170 | NA | 120 |
| Acenaphthylene | 208-96-8 | NS | 170 | NA | 120 |
| 2,6-Dinitrotoluene | 606-20-2 | NS | 170 | NA | 120 |
| 3-Nitroaniline | 99-09-2 | NS | 330 | NA | 120 |
| Acenaphthene | 83-32-9 | NS | 170 | NA | 120 |
| 2,4-Dinitrophenol | 51-28-5 | NS | 330 | NA | 800 |
| 4-Nitrophenol | 100-02-7 | NS | 330 | NA | 400 |
| Dibenzofuran | 132-64-9 | NS | 170 | NA | 120 |
| 2,4-Dinitrotoluene | 121-14-2 | NS | 170. | NA | 120 |
| 2,3,4,6-Tetrachlorophenol | 58-90-2 | NS | 170 | NA | 120 |

QAPP Worksheet #15C (Concluded) Reference Limits and Évaluation Table

Matrix:

Sediment

Analytical Group:

Semi - Volatile Organic Compounds

Concentration Level:

Medium

| Analyte | CAS Number | | | Achievable Laboratory (DESA) Limits MDLs µg/kg RLs µg/kg | | |
|-----------------------------|---------------|----|------|--|-----|--|
| Fluorene | 86-73-7 | NS | 170 | NA | 120 | |
| Diethylphthalate | 84-66-2 | NS | 170 | NA | 120 | |
| 4-Chlorophenyl Phenyl Ether | 7005-72-3 | NS | 170 | NA | 120 | |
| 4-Nitroaniline | 100-01-6 | NS | 330 | ŅA | 120 | |
| 4,6-Dinitro-2-Methylphenol | 534-52-1 | NS | 330 | NA | 400 | |
| N-Nitrosodiphenylamine | 86-30-6 | NS | 170 | NA | 120 | |
| 4-Bromophenyl Phenyl Ether | 101-55-3 | NS | 170 | NA | 120 | |
| Hexachlorobenzene | 118-74-1 | NS | 170 | NA | 120 | |
| Atrazine | 1912-24-9 | NS | 170 | NA | 120 | |
| Pentachlorophenol | 87-86-5 | NS | 330 | NA | 400 | |
| Phenanthrene | 85-01-8 | NS | 1.70 | NA. | 120 | |
| Anthracene | 120-12-7 | NS | 170 | NA | 120 | |
| Carbazole | 86-74-8 | NS | 170 | NA | 120 | |
| Di-N-Butyl Phthalate | 84-74-2 | NS | 170 | NA | 120 | |
| Fluoranthene | 206-44-0 | NS | 170 | NA | 120 | |
| Pyrene | 129-00-0 | NS | 170 | NA | 120 | |
| Butylbenzylphthalate | 85-68-7 | NS | 170 | NA | 120 | |
| 3,3-Dichlorobenzidine | 91-94-1 | NS | 170 | NA | 120 | |
| Benzo(A)Anthracene | 56-55-3 | NS | 170 | . NA | 120 | |
| Chrysene | 218-01-9 | NS | 170 | NA | 120 | |
| Bis(2-Ethylhexyl)Phthalate | 117-81-7 | NS | 170 | NA | 120 | |
| Di-N-Octyl Phthalate | 117-84-0 | NS | 170 | NA | 120 | |
| Benzo(B)Fluoranthene | 205-99-2 | NS | 170 | NA | 120 | |
| Benzo(K)Fluoranthene | 207-08-9 | NS | 170 | NA | 120 | |
| Benzo(A)Pyrene | 50-32-8 | NS | 170 | NA | 120 | |
| Indeno(1,2,3-Cd)Pyrene | 193-39-5 | NS | 170 | NA | 120 | |
| Dibenzo(A,H)Anthrcene | 53-70-6-3 | NS | 170 | NA | 120 | |
| Benzo(G,H,I)Perylene | 191-24-2 | NS | 170 | NA | 120 | |
| 1,4-Dioxane | - | NS | | NA | - | |

Note: Based on the new CRQLs the MDL study is currently being reanalyzed.

NS - Not Specified

NA - Not applicable

QAPP Worksheet #15D Reference Limits and Evaluation Table

Matrix:

Sediment

Analytical Group:

PEST/PCBs

Concentration Level:

Medium

| | CAS | Project (PRP) Quantitation | Method QLs | Achievable Laboratory (DESA) Limits | | |
|-----------------------|------------|----------------------------------|------------|--|-----------|--|
| Analyte | Number | Limit 3 | μg/kg | MDLs µg/kg | RLs µg/kg | |
| alpha-BHC | 319-89-6 | NS | 1.7 | 2.15 | 2.5 | |
| gamma-BHC | 58-89-9 | NS | 1.7 | 1.89 | 2,5 | |
| beta-BHC | 319-85-7 | NS | 1.7 | 1.35 | 2.5 | |
| delta-BHC | 319-86-8 | NS | 1.7 | 1.51 | 2.5 | |
| Heptachlor | 76-44-8 | NS | 1.7 | 2.05 | 2.5 | |
| Aldrin | 309-00-2 | NS | 1.7 | 1.66 | 2.5 | |
| Heptachlor Epoxide | 1024-57-3 | NS | 1.7 | 1.34 | 2.5 | |
| Gamma-Chlordane | 5103-74-2 | NS | 1.7 | 0.96 | 2.5 | |
| Alpha-Chlordane | 5103-71-9 | NS | 1.7 | 1.01 | 2.5 | |
| Endosulfan I | 1031-07-8 | NS | 1.7 | 1.16 | 2.5 | |
| 4,4'-DDE | 72-55-9 | NS | 3.3 | 1.92 | 5.0 | |
| Dieldrin | 60-57-1 | NS | 3.3 | 1.91 | 5.0 | |
| Endrin | 72-20-8 | NS | 3.3 | 1.84 | 5.0 | |
| 4,4'-DDD | 72-54-8 | NS | 3.3 | 1.35 | 5.0 | |
| Endosulfan Ii | 1031-078 | NS | 3.3 | 1.27 | 5.0 | |
| 4,4 ^r -DDT | 50-29-3 | NS | 3.3 | 1.52 | 5.0 | |
| Endrin Aldehyde | 7421-93-4 | ŃŚ | 3.3 | 2.24 | 5.0 | |
| Methoxychlor | 72-43-5 | NS | 17 | 8.00 | 25 | |
| Endosulfan Sulfate | 1031-07-8 | NS | 3.3 | 1.24 | 2.5 | |
| Endrin Ketone | 53494-70-5 | NS | 3.3 | 1.18 | 2.5 | |
| Toxaphene | 8001-35-2 | NS | 170 | 75.9 | 190 | |
| Technical Chlordane | | NS | NS | 56.1 | 62 | |
| AROCLOR 1016 | 12674-11-2 | NS | 33 | NS | 31 | |
| AROCLOR 1221 | 11104-28-2 | NS | 33 | NS | 62 | |
| AROCLOR 1232 | 11141-16-5 | NS | 33 | NS | 31 | |
| AROCLOR 1242 | 53469-21-9 | NS | 33 | 29.9 | 31 | |
| AROCLOR 1248 | 12672-29-6 | NS | 33 | NS | 31 | |
| AROCLOR 1254 | 11097-69-1 | NS | 33 | NS | 31 | |
| AROCLOR 1260 | 11096-82-5 | NS | 33 | NS | 31 | |
| AROCLOR 1262 | 37324-23-5 | NS | 33 | NS / | 31 | |
| AROCLOR 1268 | 11100-14-4 | NS | 33 | NS | 31 | |

NS- Not Specified

QAPP Worksheet #15E Reference Limits and Evaluation Table

Matrix:

Sediment

Analytical Group:

Metals

Concentration Level:

Medium

| | CAS | Project (PRP) Quantitation | Method CRQLs | | Laboratory) Limits |
|-----------|-----------|----------------------------------|-----------------|------------|------------------------|
| Analyte | Number | Limit 3 | mg/kg | MDLs mg/kg | RLs mg/kg |
| Aluminum | 7429-90-5 | NS | 20 | * | 100 |
| Antimony | 7440-36-0 | NS | 6 | 0.22 | 2 |
| Arsenic | 7440-38-2 | NS | 1 | 0.35 | 0.8 |
| Barium | 7440-39-3 | NS | 20 | 0.24 | 10 |
| Beryillum | 7440-41-7 | NS | 0.5 | 0.02 | 0.3 |
| Cadmium | 7440-43-9 | NS | 0.5 | 0.02 | 0,3 |
| Calcium | 7440-70-2 | NS | 500 | 12.57 | 50 |
| Chromium | 7440-47-3 | NS | 1 | 0.34 | 0.5 |
| Cobalt | 7440-48-4 | NS | 5 | 0.03 | 2 |
| Copper | 7440-50-8 | NS | 2.5 | 0.26 | 1 |
| Iron | 7439-89-6 | NS | 10 | * | 5 |
| Lead | 7439-92-1 | NS | 1 | 0.23 | 0.8 |
| Magnesium | 7439-95-4 | NS | 500 | 5.06 | - 50 |
| Manganese | 7439-96-5 | NS | 1.5 | 0.33 | 0.5 |
| Mercury | 7439-97-6 | NS | 0.1 | .0043 | 0.05 |
| Nickel | 7440-02-0 | NS | 4 | 0.09 | -2 |
| Potassium | 7440-09-7 | NS | 500 | 12.36 | 50 |
| Selenium | 7782-49-2 | NS | 3.5 | 0.22 | 2 |
| Silver | 7440-22-4 | NS | 1 | 0.06 | 0.5 |
| Sodium | 7440-23-5 | NS | 500 | 22.48 | 100 |
| Thallium | 7440-28-0 | NS | 2.5 | 3.14 | 2 |
| Vanadium | 7440-62-2 | NS | 5 | 0.40 | 2 |
| Zinc | 7440-66-6 | NS | 6 | 1.57 | 2 |
| Cyanide , | 57-12-5 | NS | NA | NA | NA |

^{*} MDL study cannot be successfully performed on these analytes because of high background levels in matrix (sand).

NS- Not Specified

QAPP Worksheet #16: Project Schedule/Timeline Table

| | | Dates (MI | M/DD/YY) | | |
|--|---|-----------------------------------|--------------------------------|----------------------|---------------------------|
| Activities | Organization | Anticipated Date(s) of Initiation | Anticipated Date of Completion | Deliverable | Deliverable Due Date |
| Preparation of QAPP | RST 2 Contractor Site Project Manager | Prior to sampling date | 4/29/2014 | QAPP | 4/30/2014 |
| Review of QAPP | RST 2 Contractor QAO and/or Group Leader | Prior to sampling date | 4/30/2014 | Approved QAPP | 5/1/2014 |
| Preparation of Health and Safety Plan | RST 2 Contractor Site Project Manager | Prior to sampling date | 4/29/2014 | HASP | 4/30/2014 |
| Procurement of Field Equipment | RST 2 Contractor Site Project Manager and/or Equipment Officer | Prior to sampling date | | NA | NA |
| Laboratory Request | RST 2 Contractor Site Project Manager and/or QAO | Prior to sampling date | 4/24/2014 | CLP Request Forms | NA |
| Field Reconnaissance/Access | RST 2 Contractor Site Project Manager, or EPA Region 2 OSC | NA | NA | NA NA | NA |
| Collection of Field Samples | RST 2 Contractor Site Project Manager | 5/5/2014 | 5/6/2014 | NA | NA |
| Laboratory Electronic Data Received | EPA Region 2 DESA | 14 Days After Sampling | 14 Days After Sampling | Preliminary Data | 14 Days After Sampling |
| Laboratory Package Received | EPA Region 2 DESA | 21 Days After Sampling | 21 Days After Sampling | Validated Data | 5/28/2014 |
| Validation of Laboratory Results | EPA Region 2 DESA | 42 Days After Sampling | 42 Days After Sampling | Final Report | 6/18/2014 |
| Data Evaluation/ Preparation of Final Report | RST 2 Contractor Site Project Manager | 6/18/2014 | 6/30/2014 | Final Report | 6/30/2014 |

QAPP Worksheet #17: Sampling Design and Rationale

RST 2 is tasked with the collection of sediment samples which will be collected from both onsite and off-site locations, including wetland areas. Sediment sampling activities will be conducted in accordance with guidelines outlined in EPA/ERT Soil Sampling SOP #2016. Sediment samples will be collected from downstream locations first and continue moving upstream so that the downstream locations are not impacted by the disturbance of sediment form upstream locations. Sediment samples will be collected at up to 20 locations utilizing disposable sampling equipment eliminating the need for equipment decontamination. TCL VOCs samples will be immediately collected utilizing Encore samplers. Then, other sediment samples will be collected with a spatula or sterile scoops and homogenized in a stainless steel bowl or pie pans. The homogenized samples will be placed in glass sample jars for TCL SVOC, PCB, pesticide, TAL Metal, including mercury and cyanide, analyses. Rocks and other debris will be removed from each sample to ensure sufficient sample volume prior to placing it into a jar. Samples must be shipped to the laboratory at the end of each day to meet the required holding time for Encore samples.

The following laboratories will provide the analyses indicated:

| Lab Name/Location | Sample Type | Parameters |
|---|-------------|--|
| EPA Region 2 DESA Laboratory 2890 Woodbridge Ave. Bldg. 209, MS-230 Edison, New Jersey 08837 | Sediment | TCL-VOC, TCL-SVOC, TAL- metals (including mercury and cyanide), PCBS, Pesticides |

Refer to Worksheet #20 for QA/QC samples, sampling methods and SOP.

QAPP Worksheet #18: Sampling Locations and Methods/SOP Requirements Table

| Matrix | Sampling Location(s) | Units | Analytical Group(s) | Concentration Level | No. of Samples (identify field duplicates) | Sampling SOP Reference | Rationale for Sampling Location |
|----------|---|-------|--|------------------------|---|------------------------------|--|
| Sediment | Approximately 5 samples per designated transect lines | mg/kg | TCL-VOCs, TCL-SVOCs, PCBs, Pesticides, and TAL Metals (including mercury & cyanide), | Low to Medium | Up to 20 (1 field duplicate) | <u>SOP#2016</u> | Characterize current sediment conditions in on-site and adjoining wetland areas. |

The website for EPA-ERT SOPs is: http://www.ert.org/mainContent.asp?section=Products&subsection=List

QAPP Worksheet #19: Analytical SOP Requirements Table

| Matrix | Analytical Group [Lab Assignment] | Concentration Level | Analytical and Preparation Method/SOP Reference | Sample Volume | Containers (number, size, and type) | Preservation Requirements | Maximum Holding Time (preparation/ analysis) |
|----------|-------------------------------------|------------------------|--|-------------------------------|-------------------------------------|--|---|
| Sediment | TCL Volatiles + Percent Moisture | Low- Medium | C-123 (<i>Ref:</i> SOM01.1) | Same(IV) | | Cool, 4°C or Frozen (-10 to - 14) | 48 hours |
| Sediment | TCL Semi-Volatiles | Low | C-90 (Ref: EPA 625) | 1 x 250g 1 x 250g(QC) | Glass, wide mouth | Cool, 4°C | To extraction: 14 days;40 days to analysis |
| Sediment | Pesticides/PCBs | Low | C-91 (Ref: EPA 608) | 1 x 100g 1 x 100g(QC) | Glass, wide mouth | Cool, 4°C | To extraction: 14 days;40 days to analysis |
| Sediment | TAL Metals/Mercury and Cyanide | Low | C-109 (Ref: EPA 200.7) C-110 (Ref: EPA 245.1) | 1 X 250ml 1 X 250ml(QC) | Rigid Plastic | Cool, 4°C | 6 months Hg- 28days CN – 14 days |

Additional volumes will be collected for MS/MSD analysis for soil matrix.

QAPP Worksheet #20: Field Quality Control Sample Summary Table

| Matrix | Analytical Group | Concentration Level | Analytical and Preparation SOP Reference | No. of Sampling Locations | No. of Field Duplicate Pairs | No. of Extra Volume Laboratory QC (e.g., MS/MSD) Samples | No. of Rinsate Blanks | No. of Trip. Blanks | Total No. of Samples to Lab |
|----------|---------------------------|------------------------|--|------------------------------------|------------------------------------|--|--------------------------|---------------------------|-----------------------------------|
| | TCL VOC | Medium | DESA C-123, C-89 | | 1/20 samples | 1/20 samples per matrix | NR | NR 1 | |
| Sediment | TCL SVOC | Medium | DESA C-90 | Approx. 5 per Transect (4 | | 1/20 samples per matrix | NR | NR | 21 |
| Scannent | TAL Metals + Mg and CN | Medium | DESA C-109, C-116, C-28 | transect (4 transects total) | per matrix | 1/20 samples per matrix | NR | NR | |
| | Pesticides/PCBS | Medium | DESA C-91 | | | 1/20 samples per matrix | NR | NR | |

Additional volume will be collected for MS/MSD Analysis. NR – Not Required

QAPP Worksheet #21: Project Sampling SOP References Table

| Reference Number | Title, Revision Date and/or Number | Originating Organization | Equipment Type | Modified for Project Work? (Y/N) | Comments |
|---------------------|--|-----------------------------|---|--|--|
| SOP#2001 | General Field Sampling Guidelines | EPA/OSWER/ERT | Flags, disposable plastic scoops, En Core volumetric sampling device | N | |
| SOP #2016 | Sediment Sampling from Compendium of ERT Surface Water and Sediment Sampling Procedures January 1991 | EPA/OSWER/ERT | Flags, disposable plastic scoops, En Core volumetric sampling device | N | 20 Sediment Samples from on-site and off-site wetlands |

See attachment B for SOPs. Note: The website for EPA-ERT SOPs is: www.ert.org/mainContent.asp?section=Products&subsection=List

QAPP Worksheet #22: Field Equipment Calibration, Maintenance, Testing, and Inspection Table

| Field Equipment | Calibration Activity | Maintenance Activity | Testing/ Inspection Activity | Frequency | Acceptance Criteria | Corrective Action | Responsible Person | SOP Reference |
|----------------------------|--|--|------------------------------------|--|--|------------------------------------|-----------------------|------------------|
| MultiRAE Plus Incl. PID | Calibrate with Zero Air; LEL: 2.5% (50% LEL) O ₂ : 18% H ₂ S: 10 ppm CO: 50 ppm PID: 100 ppm Isobutylene | Check/ replace battery/ Clean tip or bulb if necessary | Bump Test | Prior to day's activities; anytime anomaly suspected | LEL: 48-52% LEL (2% LEL) O ₂ : 17-19% (1%) H ₂ S: 9-11 ppm (1 ppm) CO: 48-52 ppm (2 ppm) PID: 95-105 ppm Isobutylene (5 ppm) | Replace battery or Replace Unit | Equipment Vendor | - |

QAPP Worksheet #23: Analytical SOP References Table

| Reference Number | Title, Revision Date, and/or Number | Definitive or Screening Data | Analytical Group | Instrument | Organization Performing Analysis | Modified for Project Work? (Y/N) |
|---------------------|---|---------------------------------|----------------------------|--------------|--|--|
| C-28 | Cyanides, Total, Rev 2.0, 3/07 | Definite | Cyanide | AutoAnalyzer | DESA LAB | N |
| C-89 | Analysis of Volatile Organic Compounds in Aqueous, Soil/Sediment and Waste Oil/Waste Organic Solvents Samples by Purge and Trap GC/MS, Rev 2.0, 3/07 | Definite | TCL Volatiles (Aqueous) | GC-MS | DESA LAB | N |
| C-123 | Analysis of Volatile Organic Compounds by Automated Closed System by Purge and Trap GC/MS, Rev 2.0, 3/07 | Definite | TCL Volatiles(Low Soil) | GC-MS | DESA LAB | N . |
| C-90 | Analysis of Base/Neutral and Acid Compounds in Aqueous, Soil/Sediment and Waste Oil/Waste Organic Solvent Samples, Rev 2.0, 3/07 | Definite | TCL Semi-Volatiles | GC-MS | DESA LAB | N |
| C-91 | Analysis of Pesticides and PCBs in Aqueous, Soil/Sediments and Waste Oil/Transformer Fluid Matrices, Rev 2.0, 3/07 | Definite | Pesticides/PCBs | GC-ECD | DESA LAB | N |
| C-109 | Determination of Trace Elements in Aqueous Trace Metals in Aqueous, Soil/Sediment/Sludge- ICP-AES, Rev 2.0, 3/07 | Definite | TAL Metals | ICP-AES | DESA LAB | N |
| C-110 | Mercury Analysis in Water and Soil/Sediments By CVAAS, Rev 2.0, 3/07 | Definite | Mercury | CVAA | DESA LAB | N |

QAPP Worksheet #24: Analytical Instrument Calibration Table

| Instrument | Calibration Procedure | Frequency of Calibration | Acceptance Criteria | Corrective Action (CA) | Person Responsible for CA | SOP Reference |
|-------------------------------|--|--------------------------|---------------------|------------------------|---------------------------|-----------------|
| ICP-AES | See SOP C-109 | See SOP C-109 | See SOP C-109 | See SOP C-109 | Assigned Lab personnel | SOP C-109 |
| CVAAS | See SOP C-110 | See SOP C-110 | See SOP C-110 | See SOP C-110 | Assigned Lab personnel | SOP C-110 |
| Colorimetric/Auto Analyzer | See SOP C-28 Per manufacture's manual | See SOP C-28 | See SOP C-28 | See SOP C-28 | Assigned Lab personnel | SOP C-28 |
| GC-ECD | See SOP C-91 | See SOP C-91 | See SOP C-91 | See SOP C-91 | Assigned Lab personnel | SOP C-91 |
| GC-MS | See SOP C- 90, C-89 | See SOP C- 90, C-89 | See SOP C- 90, C-89 | See SOP C- 90, C-89 | Assigned Lab personnel | SOP C- 90, C-89 |

QAPP Worksheet #25: Analytical Instrument and Equipment Maintenance, Testing, and Inspection Table

DESA Laboratory

| Instrument/ Equipment | Maintenance Activity | Testing Activity | Inspection Activity | Frequency | Acceptance Criteria | Corrective Action | Responsible Person | SOP Reference |
|--|-------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|
| See list of Instrument given in Worksheet #24 | See LQMP, G-10, G-11, G-12, G-19 |
| Worksheet #24 | G-11, G-12, G-19 | G-11, G-1 |

Specify the appropriate reference letter or number from the Analytical SOP References table (Worksheet #23).

QAPP Worksheet #26: Sample Handling System

SAMPLE COLLECTION, PACKAGING, AND SHIPMENT

Sample Collection (Personnel/Organization): RST 2 Site Project Manager, Weston Solutions, Inc., Region II

Sample Packaging (Personnel/Organization): RST 2 Site Project Manager and sampling team members, Weston Solutions, Inc., Region II

Coordination of Shipment (Personnel/Organization): RST 2 Site Project Manager, sampling team members, Weston Solutions, Inc., Region II

Type of Shipment/Carrier: FedEx and/ or hand-delivery.

SAMPLE RECEIPT AND ANALYSIS

Sample Receipt (Personnel/Organization):): Sample Custodian, DESA Laboratories

Sample Custody and Storage (Personnel/Organization): Sample Custodian, DESA Laboratories

Sample Preparation (Personnel/Organization): Sample Technicians, DESA Laboratory

Sample Determinative Analysis (Personnel/Organization): Sample Technicians, DESA Laboratories

SAMPLE ARCHIVING

Field Sample Storage (No. of days from sample collection): Samples to be shipped on day of collection, and arrive at laboratory within 24 hours (1 day) of sample shipment.

Sample Extract/Digestate Storage (No. of days from extraction/digestion): As per analytical methodology; see Worksheet #19.

Biological Sample Storage (No. of days from sample collection): N/A

SAMPLE DISPOSAL

Personnel/Organization: Sample Technicians, DESA Laboratories

Number of Days from Analysis: Until analysis and QA/QC checks are completed; as per analytical methodology; see Worksheet #19.

QAPP Worksheet #27: Sample Custody Requirements

Sample Identification Procedures: Each sample collected by Region II RST 2 will be identified by a property location number, the matrix of the sample collected, sample location number, the sample number, and sample type. The matrix identifier for sediment will be SD. The last number will represent the sample type (normal or duplicate). Normal samples will be identified with a 01, duplicate samples will be designated 02. The Trip blank sample will be identified by a six digit number comprised of the corresponding year, month, and day (YYMMDD).

e.g. P001-SD001-001-01 - Property 001, Sediment- Sample Location 001, sample number 001, sample type 01. TB-140501 - Trip Blank from May 1, 2014

Location of the sample collected will be recorded in the project database and site logbook. A duplicate sample will be identified in the same manner as other samples and will be distinguished and documented in the field logbook. Each sample will also be labeled with a non-CLP assigned number. Depending on the type of sample, additional information such as sampling round, date, etc. will be added.

Field Sample Custody Procedures (sample collection, packaging, shipment, and delivery to laboratory): Each sample will be individually identified and labeled after collection, then sealed with custody seals and enclosed in a plastic cooler. The sample information will be recorded on chain-of custody (COC) forms, and the samples FedEx and/or hand delivered to the appropriate laboratory by RST 2. Chain-of-custody records must be prepared in Scribe database for all CLP samples to accompany samples from the time of collection and throughout the shipping process. Each individual in possession of the samples must sign and date the sample COC Record. The chain-of-custody record will be considered completed upon receipt at the laboratory. A traffic report and chain-of-custody record will be maintained from the time the sample is taken to its final deposition. Every transfer of custody must be noted and signed for, and a copy of this record kept by each individual who has signed. When samples are not under direct control of the not under direct control of the individual responsible for them, they must be stored in a locked container sealed with a custody seal. Specific information regarding custody of the samples projected to be collected on the weekend will be noted in the field logbook.

| | | | | | | Superior Barrel and Drum Site, Revision 00 | | |
|---|---|--------------|---|---|---|--|--|--|
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Site-Specific Quality Assurance Project Plan

QAPP Worksheet #27: Sample Custody Requirements (Concluded)

The chain-of-custody record should include (at minimum) the following: 1) Sample identification number; 2) Sample information; 3) Sample location; 4) Sample date; 5) Sample Time; 6) Sample Type Matrix; 7) Sample Container Type; 8) Sample Analysis Requested; 9) Name(s) and signature(s) of sampler(s); and 10) Signature(s) of any individual(s) with custody of samples. A separate chain-of-custody form must accompany each cooler for each daily shipment. The chain-of-custody form must address all samples in that cooler, but not address samples in any other cooler. This practice maintains the chain-of-custody for all samples in case of misshipment.

Laboratory Sample Custody Procedures (receipt of samples, archiving, and disposal): A sample custodian at the laboratory will accept custody of the shipped samples, and check them for discrepancies, proper preservation, integrity, etc. If noted, issues will be forwarded to the laboratory manager for corrective action. The sample custodian will relinquish custody to the appropriate department for analysis. At this time, no samples will be archived at the laboratory. Disposal of the samples will occur only after analyses and QA/QC checks are completed.

Note: Refer to Contract Laboratory Program Guidance for Field Samplers, EPA-540-R-07-06, January 2011 at: http://www.epa.gov/superfund/programs/clp/download/sampler/clp_sampler_guidance.pdf

QAPP Worksheet # 28A QC Samples Table Volatile Organic Compounds (DESA)

| Matrix | Sediment/Aqueous ¹ | | | • | • | |
|---|---|--|---------------------------------------|---|---------------------------------|--|
| Analytical Group | VOC | | | | | |
| Concentration Level | Low/Medium | | | | | |
| Sampling SOP | ERT SOP 2016 | | | | | |
| Analytical Method/ SOP Reference | C-89/C-123 (Ref: EPA 624) | | | | | |
| Sampler's Name | Peter Lisichenko | | | | | |
| Field Sampling Organization | Weston Solutions | | | | | |
| Analytical Organization | USEPA Region 2 Lab | | | | | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | | | |
| QC Sample: | Frequency/Number | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| Tuning | 12 hr period | Pass all PBFB tune criteria | Check Instrument Reanalyze, Retune | Lab personnel | Sensitivity | Pass all PBFB tune criteria |
| Initial Calibration | SOP C-89 | % RSD +/- 35% Not more than 10% of total analytes failure % RSD not more than 60% | Check Instrument, Reanalyze | Lab personnel | Accuracy/ Precision | % RSD +/- 35% Not more than 10% of total analytes failure % RSD not more than 60% |
| Continuing Calibration Check Standard (Alternate check standard) | l per analytical batch of 20 samples | Max %D RRF +/- 30% Not more than 10% of total analytes failure % D not more than 60% | Reanalyze, Qualify data | Lab personnel | Accuracy | Max %D RRF +/- 30% Not more than 10% of total analytes failure % D not more than 60% |
| Method Blank | 1 per extraction batch of 20 samples | < RL | Investigate source of contamination | Lab personnel | Sensitivity Contamination | < RL |

QAPP Worksheet #28A (Concluded) QC Samples Table Volatile Organic Compounds (DESA)

| Matrix | Sediment/Aqueous ¹ | | | | | |
|--------------------------------|---|--|---|---|------------------------------------|---|
| Analytical Group | VOC | | | | ÷ | • |
| Concentration Level | Low/Medium | | | | • | |
| Sampling SOP | ERT SOP 2016 | | | • | | |
| Analytical Method/ SOP | C-89/C-123 | | | | | • |
| Reference | (Ref: EPA 624) | | | • | | |
| Sampler's Name | Peter Lisichenko | • | | • | | • |
| Field Sampling Organization | Weston Solutions | | • | | | |
| Analytical Organization | USEPA Region 2 Lab | | | | | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | | | |
| QC Sample: | Frequency/Number | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| Trip Blank | 1 in cooler containing first set of VOC samples | Client Defined | Investigate source of contamination | Lab personnel | Sensitivity Contamination | |
| LCS/LFB | 2 per extraction batch of 20 samples | Limits: Average Recovery 70-130% % RPD < 20 | Qualify data unless high recovery and/or Not Detected) | Lab personnel | Accuracy/ Precision | Limits: Average Recovery 70- 130% % RPD < 20 |
| Laboratory Matrix spikes | 1 per extraction batch of 20 samples | Table 4 of C-89 compound specific (full range- 17-259%) | Qualify data unless high recovery and/or Not Detected) | Lab personnel | Accuracy | Table 4 of C-89 compound specific (full range- 17-259%) |
| Internal Standards | Each sample, standard, blank | Factor of two(-50% to + 100%) from the initial/continuing calibration | Check Instrument Analyse / Qualify data | Lab personnel | Quantitation . | Factor of two(-50% to + 100%) from the initial/continuing calibration |
| Surrogates | Each sample, standard, blank | Limits 70%-130% | Reinject, Qualify data | Lab personnel | Extraction efficiency, Accuracy | Limits 70%-130% |

SOP – Sample Operating Procedure

1 Aqueous sample will consist of trip blank sample only. Aqueous field duplicate and MS/MSD will not be collected

QAPP Worksheet #28B

QC Samples Table TCL Semivolatile Organic Compounds (DESA)

| | | == | , | | | • |
|---|---|--|---------------------------------------|---|---------------------------------|--|
| Matrix | Sediment |] | | | • | |
| Analytical Group | SVOC | | | | | |
| Concentration Level | Medium | | | • | · | |
| Sampling SOP | ERT SOP 2016 | | | | | |
| Analytical Method/ SOP Reference | C-90 (Ref: EPA 625) | | | | | |
| Sampler's Name | Peter Lisichenko | ` | | | | |
| Field Sampling Organization | Weston Solutions | | | | | |
| Analytical Organization | USEPA Region 2 Lab | | | | | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | | | |
| QC Sample: | Frequency/Number | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| Tuning | 12 hr period | Pass all DFTPP tune criteria | Check Instrument Reanalyze, Retune | Lab personnel | Sensitivity | Pass all DFTPP tune criteria |
| Initial Calibration | SOP C-90 | % RSD +/- 35% Allowed to fail 10% of total number of analytes but % RSD not be more than 60% | Check Instrument, Reanalyze | Lab personnel | Accuracy/ Precision | % RSD +/- 35% Allowed to fail 10% of total number of analytes but % RSD not be more than 60% |
| Continuing Calibration Check Standard (Alternate check standard) | l per analytical batch of ≤20 samples | Min RRF 0.05 Max %D +/- 20% 10% of total analytes allowed to fail but not more than 60% | Reanalyze, Qualify data | Lab personnel | Accuracy | Min RRF 0.05 Max %D RRF +/- 20% 10% of total analytes allowed to fail but not more than 60% |
| Method Blank | 1 per extraction batch of ≤20 samples | < RL | Investigate source of contamination | Lab personnel | Sensitivity Contamination | < RL |

QAPP Worksheet #28B (Concluded) QC Samples Table TCL Semivolatile Organic Compounds (DESA)

| | | -n | | | | |
|-------------------------------------|--|---|---|---|---------------------------------|--|
| Matrix | Sediment | | | | | |
| Analytical Group | SVOC | | | • | | |
| Concentration Level | Medium | | | 4 | | |
| Sampling SOP | ERT SOP 2016 | | • | | • | |
| Analytical Method/ SOP Reference | C-90 (Ref: EPA 625) | | | | | |
| Sampler's Name | Peter Lisichenko | | | | | |
| Field Sampling Organization | Weston Solutions | | | | | |
| Analytical Organization | USEPA Region 2 Lab | | | | • | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | • | | |
| QC Sample: | Frequency/Number | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| LCS/LFB | 2 per extraction batch of ≤20 samples | Limits listed in Table3 in SOP C-90 for aqueous, manufacture's limits for soil % RPD < 30 | Qualify data unless high recovery and/or Not Detected) | Lab personnel | Accuracy/ Precision | Limits listed in Table3 in SOP C-90 for aqueous, manufacture's limits for soil % RPD < 30 |
| Laboratory Matrix spikes | 1 per extraction batch of ≤20 samples | Limits listed in Table3 in SOP C-90 | Qualify data unless high recovery and/or Not Detected) | Lab personnel | Accuracy | Limits listed in Table3 in SOP C-90 |
| Internal Standards | Each sample, standard, blank | Factor of two (-50% to -100%) | Check Instrument Analyse / Qualify data | Lab personnel | Quantitation | Factor of two (-50% to -100%) |
| Surrogates | Each sample, standard, blank | 30%-120% for Base Neutrals 20-120% for Acids | Reinject, Qualify data as per SOP C-90 | Lab personnel | Extraction efficiency, Accuracy | 30%-120% for Base Neutrals 20-120% for Acids |

SOP - Sample Operating Procedure

QAPP Worksheet #28C QC Samples Table TCL PCBs and Pesticides (DESA)

| G | 1 | 1 | | | • | |
|--------------------------------|---|--|--|---|---------------------------------|--|
| Matrix | Sediment | | | | | · |
| Analytical Group | Pesticides/PCBs | · | | • | | |
| Concentration Level | Medium | | | | | |
| Sampling SOP | ERT SOP 2016 | | | | | |
| Analytical Method/ SOP | C-91 | | | | | |
| Reference | (Ref: EPA 608) | | | | • | |
| Sampler's Name | Peter Lisichenko | · | | • | | |
| Field Sampling Organization | Weston Solutions | | | | | |
| Analytical Organization | USEPA Region 2 Lab | | | | • | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | | | |
| QC Sample: | Frequency/Number | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| Instrument | D | Total breakdown | | | 0 1.1.1. | Total breakdown |
| Performance (PEM) | Beginning of each analytical run | otal breakdown <30% | Check Instrument | Lab personnel | Sensitivity Contamination | <30% |
| | | | Check Instrument Check Instrument, Reanalyze | Lab personnel Lab personnel | | |
| Performance (PEM) | analytical run C-91 | <30% % RSD +/- 25% Not more than 10% of total analytes failure RSD not more than | Check Instrument, | • | Contamination Accuracy/ | <30% % RSD +/- 25% Not more than 10% of total analytes failure |

QAPP Worksheet #28C (Concluded) QC Samples Table TCL PCBs and Pesticides (DESA)

| Matrix | Sediment | · | • | | | |
|-------------------------------------|---|--|---|---|---------------------------------|---|
| Analytical Group | Pesticides/PCBs | | | | | |
| Concentration Level | Medium | | | | | |
| Sampling SOP | ERT SOP 2016 | | | | | |
| Analytical Method/ SOP Reference | C-91 (Ref: EPA 608) | | | • | | |
| Sampler's Name | Peter Lisichenko | | | | | • |
| Field Sampling Organization | Weston Solutions | | | | | |
| Analytical Organization | USEPA Region 2 Lab | | | | | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | | | |
| (| | | | | | |
| QC Sample: | Frequency/Number | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| QC Sample: LCS/LFB | Frequency/Number 2 per extraction batch | _ | Corrective Action Qualify data unless high recovery and/or Not Detected) | Responsible for | | Criteria Limits: Average Recovery 50- 150% % RPD < 30 |
| | | Acceptance Limits Limits: Average Recovery 50-150% | Qualify data unless high recovery and/or Not | Responsible for Corrective Action | Indicator (DQI) Accuracy/ | Criteria Limits: Average Recovery 50- 150% |

SOP - Sample Operating Procedure.

QAPP Worksheet #28D QC Samples Table TAL Metals including Mercury and Cyanide (DESA)

| Matrix | Sediment | | | • | | |
|---|---|--------------------------------------|--|---|------------------------------|---|
| Analytical Group | Metals + Mercury and Cyanide | | | ÷ | | |
| Concentration Level | Low/Medium | | ٠. | | | |
| Sampling SOP | 2016 | · | | | | |
| Analytical Method/ SOP Reference | C-109,C-112, C-110, C-28 (Ref: EPA 200.7, 200.8, 245.1) | | | | | |
| Sampler's Name | Peter Lisichenko | | | | | |
| Field Sampling Organization | Weston Solutions | | | | | |
| Analytical Organization | USEPA Region 2 Lab | | | | | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | | | |
| QC Sample: | European en Dinashon | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| 1 OC Sample: | rrequency/Number | Acceptance Linus | COLLECTIVE ACTION | Corrective Action | Indicator (DQI) | Citteria |
| Tuning/System Stability(ICP-MS) | As per C-112 | Pass all the tune/stability criteria | Check Instrument Reanalyze, Retune | Lab personnel | Sensitivity | Pass all the tune/stability criteria |
| Tuning/System | | Pass all the | Check Instrument | | | Pass all the tune/stability criteria 90%-110% |
| Tuning/System Stability(ICP-MS) Initial Calibration | As per C-112 Immediately following each calibration ,after every 10 samples and at the end of each | Pass all the tune/stability criteria | Check Instrument Reanalyze, Retune Check Instrument, | Lab personnel | Sensitivity | Pass all the tune/stability criteria |

QAPP Worksheet #28D (Continued) QC Samples Table TAL Metals including Mercury and Cyanide

| | · | ਜ | | | | |
|--|--|--|-------------------------------------|---|---------------------------------------|--|
| Matrix | Sediment |]. | | | | |
| Analytical Group | Metals + Mercury and Cyanide | | | • | | |
| Concentration Level | Low/Medium | | | • | | |
| Sampling SOP | 2016 | 1 | | | | |
| Analytical Method/ SOP Reference | C-109,C-112, C-110, C-28 (Ref: EPA 200.7, 200.8, 245.1) | | | | | |
| Sampler's Name | Peter Lisichenko | 1 | | | | |
| Field Sampling Organization | Weston Solutions | | | · • | | |
| Analytical Organization | USEPA Region 2 Lab | | | | | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | | · · · · · · · · · · · · · · · · · · · | |
| QC Sample: | Frequency/Number | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| Continuing Calibration Blank(CCB) | After every CCV | < RL | Investigate source of contamination | Lab personnel | Sensitivity Contamination | < RL |
| Low Level Check Standard | At Beginning and end of each analytical run | ± 30% of the true value | Check Instrument, Re-calibrate | Lab personnel | Accuracy | ± 30% of the true value |
| Interference Check Sample(ICP-200.7) | At Beginning and end of each analytical run | < RL Except Al ,Fe, Ca, K, Mg and Na | As per C-109 | Lab personnel | Precision | < RL Except Al ,Fe, Ca, K, Mg and Na |
| Method blank | 1 per extraction batch of ≤20 samples | < RL | Investigate source of contamination | Lab personnel | Sensitivity Contamination | < RL |
| LCS/LFB | 2 per extraction batch of ≤20 samples | Limits: Average Recovery ± 20% aqueous, ± 25% Soil) % RPD < 20(Aq), % RPD <25(Soil) | Qualify data | Lab personnel | Accuracy/ Precision | Limits: Average Recovery ± 20% aqueous, ± 25% Solids) % RPD < 20(Aq), % RPD <25(Soil |

QAPP Worksheet #28D (Concluded) QC Samples Table TAL Metals including Mercury and Cyanide

| Matrix | Sediment | 1 | | | | |
|-------------------------------------|--|---|---------------------------------------|---|---------------------------------|--|
| Analytical Group | Metals + Mercury and Cyanide | | | | | • |
| Concentration Level | Low/Medium | | • | | | |
| Sampling SOP | 2016 | 1 | | | | |
| Analytical Method/ SOP Reference | C-109,C-112, C-110, C-28 (Ref: EPA 200.7, 200.8, 245.1) | | | | | |
| Sampler's Name | Peter Lisichenko | : | | | | |
| Field Sampling Organization | Weston Solutions | ~ | | | | |
| Analytical Organization | USEPA Region 2 Lab | | | | | |
| No. of Sample Locations | Approx. 5 per Transect (4 Transects total) | | | | | |
| QC Sample: | Frequency/Number | Method/SOP QC Acceptance Limits | Corrective Action | Person(s) Responsible for Corrective Action | Data Quality Indicator (DQI) | Measurement Performance Criteria |
| Laboratory Matrix spikes | 1 per extraction batch of ≤20 samples | Limits ± 20% aqueous, ± 25% Soil) | Qualify data | Lab personnel | Accuracy | Limits ± 20% aqueous, ± 25% Soil) |
| Serial Dilution Test(ICP-200.7) | Matrix spike sample | RPD < 20 % | Qualify data | Lab personnel | Precision | RPD < 20 % |
| Internal | Each sample, standard, blank | Range of 0.60-1.87 of the original response | Check Instrument Analyse / Qualify | Lab personnel | Quantitation | Range of 0.60-1.87 of the original response in the |

SOP - Sample Operating Procedure.

QAPP Worksheet #29: Project Documents and Records Table

| Sample Collection Documents and Records | Analysis Documents and Records | Data Assessment Documents and Records | Other |
|---|--|--|--------------------|
| Site and field logbooks COC forms Field Data Sheets GIS map for sampling locations Incident Action plan | Sample receipt logs Internal and external COC forms Equipment calibration logs Sample preparation worksheets/logs Sample analysis worksheets/run logs Telephone/email logs Corrective action documentation | Data validation reports Field inspection checklist(s) Laboratory Audit checklist (if performed) Review forms for electronic entry of data into database Corrective action documentation Laboratory Final Data | • CLP Request Form |

QAPP Worksheet #30: Analytical Services Table

| Matrix | Analytical Group | Concentration Level | Analytical SOP | Data Package Turnaround Time | Laboratory/Organization (Name and Address, Contact Person and Telephone Number) | Backup Laboratory/Organization (Name and Address, Contact Person and Telephone Number) |
|----------|--------------------------|------------------------|----------------------------|------------------------------------|---|--|
| | TCL VOCs | Medium | DESA C-123 | 3 weeks written | | NA |
| | TCL SVOCs | Medium | DESA C-90 | 3 weeks written | EPA Region 2 DESA Laboratory 2890 Woodbridge Ave. | NA |
| Sediment | TAL Metals, Hg and CN | Medium | DESA C 109, C-110, C-28 | 3 weeks written | Bldg. 209, MS-230 Edison, New Jersey 08837 | NA |
| | Pesticides/PCBs | Medium | DESA C-91 | 3 weeks written | John Birri 732-906-6886 | NA . |

Note: Aqueous Trip Blank Sample will be collected for TCL VOC Analysis Only.

QAPP Worksheet #31: Planned Project Assessments Table

| Assessment Type | Frequency | Internal or External | Organization Performing Assessment | Person(s) Responsible for Performing Assessment (Title and Organizational Affiliation) | Person(s) Responsible for Responding to Assessment Findings (Title and Organizational Affiliation) | Person(s) Responsible for Identifying and Implementing Corrective Actions (CA) (Title and Organizational Affiliation) | Person(s) Responsible for Monitoring Effectiveness of CA (Title and Organizational Affiliation) |
|-------------------|-----------------|-------------------------|--|--|---|---|---|
| РТ | Semiannually | External | NELAC | PT provider | Lab Personnel | Lab Personnel | Lab QA Officer |
| NELAC | Every two years | External | NELAC | Florida DOH | Lab QA Officer | Lab Personnel | Florida DOH |
| INTERNAL AUDIT | Monthly | Internally | DESA Lab | Lab QA Officer | Lab Personnel | Lab Personnel | Lab QA Officer |

QAPP Worksheet #32 Assessment Findings and Corrective Action Responses

| Assessment Type | Nature of Deficiencies Documentation | Individual(s) Notified of Findings (Name, Title, Organization) | Timeframe of Notification | Nature of Corrective Action Response Documentation | Individual(s) Receiving Corrective Action Response (Name, Title, Org.) | Timeframe for Response |
|--|--|--|---|--|--|--|
| Project Readiness Review | Checklist or logbook entry summary | Site Project Manager, Weston Solutions, Inc. | Immediately to within 24 hours of review | Checklist or logbook entry | Site Project Manager, Weston Solutions, Inc. | Immediately to within 24 hours of review |
| Field Observations/ Deviations from Work Plan | Logbook | Site Project Manager, Weston Solutions, Inc. and EPA RPM | Immediately to within 24 hours of deviation | Logbook | Site Project Manager, Weston Solutions, Inc. and EPA RPM | Immediately to within 24 hours of deviation |
| Laboratory Technical Systems/ Performance Audits | Written Report | DESA Laboratory | 30 days | Letter | DESA Laboratory | 14 days |
| On-Site Field Inspection | Written Report | Site Project Manager, Weston Solutions, Inc. | 7 calendar days after completion of the audit | Letter/Internal Memorandum | Site Project Manager, Weston Solutions, Inc. and/or EPA RPM | To be identified in the cover letter of the report |
| Performance Evaluation Samples | Electronic Report | DESA Laboratory | 30 days | Letter or Written Report | DESA Laboratory | 14 days |
| Peer Review | Deliverables | SPM, Weston Solutions, Inc. | Prior to deliverable due date | Comments directly on deliverable | SPM, Weston Solutions, Inc. | Prior to deliverable due date |

QAPP Worksheet #33 QA Management Reports Table

| Type of Report | Frequency (daily, weekly, monthly, quarterly, annually, etc.) | Projected Delivery Date(s) | Person(s) Responsible for Report Preparation (Title and Organizational Affiliation) | Report Recipient(s) (Title and Organizational Affiliation) |
|------------------------------------|---|--|---|---|
| EPA-DESA Laboratory | As performed | Up to 60 days after receipt of unvalidated data | EPA Region 2 | Keith Glenn, EPA OSC, Region 2 and Site Project Manager, Weston Solutions, Inc. |
| EPA-DESA Laboratory (preliminary) | As performed | 2 weeks from the sampling date | EPA DESA Laboratory | Keith Glenn, EPA OSC, Region 2 and Site Project Manager, Weston Solutions, Inc. |
| EPA-DESA Laboratory (validated) | As performed | Up to 21 days after receipt of preliminary data | EPA Region II DESA & ESAT Data Validators | RST 2 SPM, Weston Solutions, Inc., and OSC, EPA Region II |
| On-Site Field Inspection | As performed | 7 calendar days after completion of the inspection | RST 2 HSO | RST 2 SPM, Weston Solutions, Inc. |
| Field Change Request | As required per field change | 3 days after identification of need for field change | RST 2 SPM | Keith Glenn EPA OSC |
| Final Report | As performed | 2 weeks after receipt of EPA approval of data package | RST 2 SPM | Keith Glenn EPA OSC |

QAPP Worksheet #34 Verification (Step I) Process Table

| Verification Input | Description | | ¹ Responsible for Verification (Name, Organization) | |
|------------------------------------|--|---|--|--|
| Site/field logbooks | Field notes will be prepared daily by the RST 2 Site Project Manager and will be complete, appropriate, legible and pertinent. Upon completion of field work, logbooks will be placed in the project files. | I | Site Project Manager, Weston Solutions, Inc. | |
| Chains of custody | COC forms will be reviewed against the samples packed in the specific cooler prior to shipment. The reviewer will initial the form. An original COC will be sent with the samples to the laboratory, while copies are retained for (1) the Sampling Trip Report and (2) the project files. | I | Site Project Manager, Weston Solutions, Inc. | |
| Sampling Trip Reports | STRs will be prepared for each week of field sampling [for which samples are sent to EPA's DESA laboratory.] Information in the STR will be reviewed against the COC forms, and potential discrepancies will be discussed with field personnel to verify locations, dates, etc. | I | Site Project Manager, Weston Solutions, Inc. | |
| Laboratory analytical data package | Data packages will be reviewed/verified internally by the laboratory performing the work for completeness and technical accuracy prior to submittal. | Е | EPA DESA Laboratory | |
| Laboratory analytical data package | Data packages will be reviewed as to content and sample information upon receipt by EPA. | I | DESA Data Validation Personnel | |
| Final Sample Report | The project data results will be compiled in a sample report for the project. Entries will be reviewed/verified against hardcopy information. | I | Site Project Manager, Weston Solutions, Inc. | |

Responsible for verifications, and their name and organization will be added

QAPP Worksheet #34 Verification (Step I) Process Table (Concluded)

| Verification Input | Description | Internal/ External | Responsible for Verification (Name, Organization) |
|--|---|-----------------------|--|
| EPA DESA Laboratory | | | |
| Chain of Custody | Chain-of-custody forms will be verified against the sample cooler they represent. Sample Acceptance Checklist is completed. The OSCAR staff supervisor utilizes the analyses request information and the external COC to review the accuracy and completeness of LIMS log-in entries, as reflected on the LIMS Sample Receipt Form Details can be found in Laboratory Quality Management Plan, SOP G-25 | | OSCAR Personnel DESA LAB |
| Analytical data package/ Final Report | The procedures for data review: 1- Data reduction/review by Primary Analyst. 2- Review complete data package (raw data) by independent Peer Reviewer 3- The Sample Project Coordinator reviews the project documentation for completeness followed by a QA review by the QAO 4- Final review by Branch Chief/Section Chief prior to release, this review is to ensure completeness and general compliance with the objectives of the project. This final review typically does not include a review of raw data. Details can be found in Laboratory Quality Management Plan. | I | Primary Analyst, Peer Reviewer, Sample Project Coordinator, Quality Assurance Officer, Section Chief/ Branch Chief. DESA LAB |

QAPP Worksheet #35 Validation (Steps IIa and IIb) Process Table

| Step IIa/IIb Validation Input | | Description | Responsible for Validation (Name, Organization) | |
|-------------------------------|---------------------------------------|---|--|--|
| IIa | SOPs | Ensure that the sampling methods/procedures outlined in QAPP were followed, and that any deviations were noted/approved. | RST 2 Site Project Manager | |
| IIb | SOPs | Determine potential impacts from noted/approved deviations, in regard to PQOs. | RST 2 Site Project Manager | |
| EPA DESA Lab | oratory | | | |
| | Chain of Custody | Chain-of-custody forms will be verified against the sample cooler they represent. Sample Acceptance Checklist is completed. The OSCAR staff supervisor utilizes the analyses request information and the external COC to review the accuracy and completeness of LIMS log-in entries, as reflected on the LIMS Sample Receipt Form Details can be found in Laboratory Quality Management Plan, SOP G-25 | OSCAR Personnel DESA LAB | |
| | Analytical data package/ Final Report | The procedures for data review: 1- Data reduction/review by Primary Analyst. 2- Review complete data package (raw data) by independent Peer Reviewer 3- The Sample Project Coordinator reviews the project documentation for completeness followed by a QA review by the QAO 4- Final review by Branch Chief/Section Chief prior to release, this review is to ensure completeness and general compliance with the objectives of the project. This final review typically does not include a review of raw data. Details can be found in Laboratory Quality Management Plan. | Primary Analyst, Peer Reviewer, Sample Project Coordinator, Quality Assurance Officer, Section Chief/ Branch Chief. DESA LAB | |
| IIa | Chains of custody | Examine COC forms against QAPP and laboratory contract requirements (e.g., analytical methods, sample identification, etc.). | DESA Data Validation Personnel | |
| IIa | Laboratory data package | Examine packages against QAPP and laboratory contract requirements, and against COC forms (e.g., holding times, sample handling, analytical methods, sample identification, data qualifiers, QC samples, etc.). | DESA Data Validation Personnel | |
| IIb | Laboratory data package | Determine potential impacts from noted/approved deviations, in regard to PQOs. Examples include PQLs and QC sample limits (precision/accuracy). | DESA Data Validation Personnel | |
| IIb | Field duplicates | Compare results of field duplicate (or replicate) analyses with RPD criteria | DESA Data Validation Personnel | |

QAPP Worksheet #36: Validation (Steps IIa and IIb) Summary Table

| Step IIa/IIb | Matrix | Analytical Group | Concentration Level | Validation Criteria | Data Validator (title and organizational affiliation) |
|--------------|----------|-------------------------------------|---------------------|--|---|
| IIa / IIb | Sediment | TCL VOCs | Low | DESA Data Method and SOP # G-26 Guideline for Laboratory Data Review | DESA Data Validation Personnel, EPA Region 2 |
| Ila / Ilb | Sediment | TCL SVOCs | Low/Medium | DESA Data Method and SOP # G-26 Guideline for Laboratory Data Review | DESA Data Validation Personnel, EPA Region 2 |
| IIa / IIb | Sediment | TAL Metals including Hg and Cyanide | Low/Medium | DESA Data Method and SOP # G-26 Guideline for Laboratory Data Review | DESA Data Validation Personnel, EPA Region 2 |
| IIa / IIb | Sediment | TCL PCBs | Low/Medium | DESA Data Method and SOP # G-26 Guideline for Laboratory Data Review | DESA Data Validation Personnel, EPA Region 2 |
| IIa / IIb | Sediment | TCL Pesticides | Low/Medium | DESA Data Method and SOP # G-26 Guideline for Laboratory Data Review | DESA Data Validation Personnel, EPA Region 2 |

QAPP Worksheet #37: Usability Assessment

Summarize the usability assessment process and all procedures, including interim steps and any statistics, equations, and computer algorithms that will be used: Data, whether generated in the field or by the laboratory, are tabulated and reviewed for Precision, Accuracy, Representativeness, Completeness, and Comparability (PARCCS) by the SPM for field data or the data validator for laboratory data. The review of the PARCC Data Quality Indicators (DQI) will compare with the DQO detailed in the site-specific QAPP, the analytical methods used and impact of any qualitative and quantitative trends will be examined to determine if bias exists. A hard copy of field data is maintained in a designated field or site logbook. Laboratory data packages are validated, and final data reports are generated. All documents and logbooks are assigned unique and specific control numbers to allow tracking and management.

Where applicable, the following documents will be followed to evaluate data for fitness in decision making: EPA QA/G-4, <u>Guidance on Systematic Planning using the Data Quality Objectives Process</u>, EPA/240/B-06/001, February 2006, and EPA QA/G-9R, <u>Guidance for Data Quality Assessment</u>, A reviewer's <u>Guide EPA/240/B-06/002</u>, February 2006.

Describe the evaluative procedures used to assess overall measurement error associated with the project:

As delineated in the Uniform Federal Policy for Implementing Environmental Quality Systems: Evaluating, Assessing and Documenting Environmental Data Collection and Use Programs Part 1: UFP-QAPP (EPA-505-B-04-900A, March 2005); Part 2A: UFP-QAPP Workbook (EPA-505-B-04-900C, March 2005); Part 2B: Quality Assurance/Quality Control Compendium: Non-Time Critical QA/QC Activities (EPA-505-B-04-900B, March 2005); "Graded Approach" will be implemented for data collection activities that are either exploratory or where specific decisions cannot be identified, since this guidance indicates that the formal DQO process is not necessary.

QAPP Worksheet #37: Usability Assessment- (Concluded)

Weston Solutions, Inc., RST 2 will conduct sediment sampling as part of the Removal Assessment of the Site. The analytical data from this investigation will be used to assist the EPA in determining the presence and nature of the contaminants at the Site and the adjoining Freshwater Forested/Shrub Wetlands.

Identify the personnel responsible for performing the usability assessment: Site Project Management Team, Data Validation Personnel, and EPA Region 2 OSC

Describe the documentation that will be generated during usability assessment and how usability assessment results will be presented so that they identify trends, relationships (correlations), and anomalies:

A copy of the most current approved QAPP, including any graphs, maps and text reports developed will be provided to all personnel identified on the distribution list.

Attachment A

Figure 1 – Site Location Map



Attachment B

EPA/ERT Standard Operating Procedures (SOPs)

SOP No.: 2001 - General Field Sampling Guidelines SOP No.: 2016 - Sediment Sampling



GENERAL FIELD SAMPLING GUIDELINES

SOP#: 2001 DATE: 08/11/94

REV. #: 0.0

1.0 SCOPE AND APPLICATION

The purpose of this Standard Operating Procedure (SOP) is to provide general field sampling guidelines that will assist REAC personnel in choosing sampling strategies, location, and frequency for proper assessment of site characteristics. This SOP is applicable to all field activities that involve sampling.

These are standard (i.e., typically applicable) operating procedures which may be varied or changed as required, dependent on site conditions, equipment limitations or limitations imposed by the procedure. In all instances, the ultimate procedures employed should be documented and associated with the final report.

Mention of trade names or commercial products does not constitute U.S. EPA endorsement or recommendation for use.

2.0 METHOD SUMMARY

Sampling is the selection of a representative portion of a larger population, universe, or body. Through examination of a sample, the characteristics of the larger body from which the sample was drawn can be inferred. In this manner, sampling can be a valuable tool for determining the presence, type, and extent of contamination by hazardous substances in the environment.

The primary objective of all sampling activities is to characterize a hazardous waste site accurately so that its impact on human health and the environment can be properly evaluated. It is only through sampling and analysis that site hazards can be measured and the job of cleanup and restoration can be accomplished effectively with minimal risk. The sampling itself must be conducted so that every sample collected retains its original physical form and chemical composition. In this way, sample integrity is insured, quality assurance standards are maintained, and the sample can accurately represent the larger body of

material under investigation.

The extent to which valid inferences can be drawn from a sample depends on the degree to which the sampling effort conforms to the project's objectives. For example, as few as one sample may produce adequate, technically valid data to address the project's objectives. Meeting the project's objectives requires thorough planning of sampling activities, and implementation of the most appropriate sampling and analytical procedures. These issues will be discussed in this procedure.

3.0 SAMPLE PRESERVATION, CONTAINERS, HANDLING, AND STORAGE

The amount of sample to be collected, and the proper sample container type (i.e., glass, plastic), chemical preservation, and storage requirements are dependent on the matrix being sampled and the parameter(s) of interest. Sample preservation, containers, handling, and storage for air and waste samples are discussed in the specific SOPs for air and waste sampling techniques.

4.0 INTERFERENCES AND POTENTIAL PROBLEMS

The nature of the object or materials being sampled may be a potential problem to the sampler. If a material is homogeneous, it will generally have a uniform composition throughout. In this case, any sample increment can be considered representative of the material. On the other hand, heterogeneous samples present problems to the sampler because of changes in the material over distance, both laterally and vertically.

Samples of hazardous materials may pose a safety threat to both field and laboratory personnel. Proper health and safety precautions should be implemented when handling this type of sample. Environmental conditions, weather conditions, or non-target chemicals may cause problems and/or interferences when performing sampling activities or when sampling for a specific parameter. Refer to the specific SOPs for sampling techniques.

5.0 EQUIPMENT/APPARATUS

The equipment/apparatus required to collect samples must be determined on a site specific basis. Due to the wide variety of sampling equipment available, refer to the specific SOPs for sampling techniques which include lists of the equipment/apparatus required for sampling.

6.0 REAGENTS

Reagents may be utilized for preservation of samples and for decontamination of sampling equipment. The preservatives required are specified by the analysis to be performed. Decontamination solutions are specified in ERT SOP #2006, Sampling Equipment Decontamination.

7.0 PROCEDURE

7.1 Types of Samples

In relation to the media to be sampled, two basic types of samples can be considered: the environmental sample and the hazardous sample.

Environmental samples are those collected from streams, ponds, lakes, wells, and are off-site samples that are not expected to be contaminated with hazardous materials. They usually do not require the special handling procedures typically used for concentrated wastes. However, in certain instances, environmental samples can contain elevated concentrations of pollutants and in such cases would have to be handled as hazardous samples.

Hazardous or concentrated samples are those collected from drums, tanks, lagoons, pits, waste piles, fresh spills, or areas previously identified as contaminated, and require special handling procedures because of their potential toxicity or hazard. These samples can be further subdivided based on their degree of hazard; however, care should be taken when handling and shipping any wastes believed to be concentrated regardless of the degree.

The importance of making the distinction between environmental and hazardous samples is two-fold:

- (1) Personnel safety requirements: Any sample thought to contain enough hazardous materials to pose a safety threat should be designated as hazardous and handled in a manner which ensures the safety of both field and laboratory personnel.
- (2) Transportation requirements: Hazardous samples must be packaged, labeled, and shipped according to the International Air Transport Association (IATA) Dangerous Goods Regulations or Department of Transportation (DOT) regulations and U.S. EPA guidelines.

7.2 Sample Collection Techniques

In general, two basic types of sample collection techniques are recognized, both of which can be used for either environmental or hazardous samples.

Grab Samples

A grab sample is defined as a discrete aliquot representative of a specific location at a given point in time. The sample is collected all at once at one particular point in the sample medium. The representativeness of such samples is defined by the nature of the materials being sampled. In general, as sources vary over time and distance, the representativeness of grab samples will decrease.

Composite Samples

Composites are nondiscrete samples composed of more than one specific aliquot collected at various sampling locations and/or different points in time. Analysis of this type of sample produces an average value and can in certain instances be used as an alternative to analyzing a number of individual grab samples and calculating an average value. It should be noted, however, that compositing can mask problems by diluting isolated concentrations of some hazardous compounds below detection limits.

Compositing is often used for environmental samples and may be used for hazardous samples under certain conditions. For example, compositing of hazardous waste is often performed after compatibility tests have been completed to determine an average value over a number of different locations (group of drums). This procedure generates data that can be useful by providing an average concentration within a number of units, can serve to keep analytical costs down, and can provide information useful to transporters and waste disposal operations.

For sampling situations involving hazardous wastes, grab sampling techniques are generally preferred because grab sampling minimizes the amount of time sampling personnel must be in contact with the wastes, reduces risks associated with compositing unknowns, and eliminates chemical changes that might occur due to compositing.

7.3 Types of Sampling Strategies

The number of samples that should be collected and analyzed depends on the objective of the investigation. There are three basic sampling strategies: random, systematic, and judgmental sampling.

Random sampling involves collection of samples in a nonsystematic fashion from the entire site or a specific portion of a site. Systematic sampling involves collection of samples based on a grid or a pattern which has been previously established. When judgmental sampling is performed, samples are collected only from the portion(s) of the site most likely to be contaminated. Often, a combination of these strategies is the best approach depending on the type of the suspected/known contamination, the uniformity and size of the site, the level/type of information desired, etc.

7.4 QA Work Plans (QAWP)

A QAWP is required when it becomes evident that a field investigation is necessary. It should be initiated in conjunction with, or immediately following, notification of the field investigation. This plan should be clear and concise and should detail the following basic components, with regard to sampling activities:

- C Objective and purpose of the investigation.
- Basis upon which data will be evaluated.
- Information known about the site including location, type and size of the facility, and length of operations/abandonment.
- C Type and volume of contaminated material, contaminants of concern (including

- concentration), and basis of the information/data.
- C Technical approach including media/matrix to be sampled, sampling equipment to be used, sample equipment decontamination (if necessary), sampling design and rationale, and SOPs or description of the procedure to be implemented.
- Project management and reporting, schedule, project organization and responsibilities, manpower and cost projections, and required deliverables.
- QA objectives and protocols including tables summarizing field sampling and QA/QC analysis and objectives.

Note that this list of QAWP components is not allinclusive and that additional elements may be added or altered depending on the specific requirements of the field investigation. It should also be recognized that although a detailed OAWP is quite important, it may be impractical in some instances. Emergency responses and accidental spills are prime examples of such instances where time might prohibit the development of site-specific QAWPs prior to field activities. In such cases, investigators would have to rely on general guidelines and personal judgment, and the sampling or response plans might simply be a strategy based on preliminary information and finalized on site. In any event, a plan of action should be developed, no matter how concise or informal, to aid investigators in maintaining a logical and consistent order to the implementation of their task.

7.5 Legal Implications

The data derived from sampling activities are often introduced as critical evidence during litigation of a hazardous waste site cleanup. Legal issues in which sampling data are important may include cleanup cost recovery, identification of pollution sources and responsible parties, and technical validation of remedial design methodologies. Because of the potential for involvement in legal actions, strict adherence to technical and administrative SOPs is essential during both the development and implementation of sampling activities.

Technically valid sampling begins with thorough planning and continues through the sample collection and analytical procedures. Administrative requirements involve thorough, accurate

documentation of all sampling activities. Documentation requirements include maintenance of a chain of custody, as well as accurate records of field activities and analytical instructions. Failure to observe these procedures fully and consistently may result in data that are questionable, invalid and non-defensible in court, and the consequent loss of enforcement proceedings.

8.0 CALCULATIONS

Refer to the specific SOPs for any calculations which are associated with sampling techniques.

9.0 QUALITY ASSURANCE/ QUALITY CONTROL

Refer to the specific SOPs for the type and frequency of QA/QC samples to be analyzed, the acceptance criteria for the QA/QC samples, and any other QA/QC activities which are associated with sampling techniques.

10.0 DATA VALIDATION

Refer to the specific SOPs for data validation activities that are associated with sampling techniques.

11.0 HEALTH AND SAFETY

When working with potentially hazardous materials, follow U.S. EPA, OSHA, and corporate health and safety procedures.



SEDIMENT SAMPLING

SOP#: 2016 DATE: 11/17/94

REV. #: 0.0

1.0 SCOPE AND APPLICATION

This standard operating procedure (SOP) is applicable to the collection of representative sediment samples. Analysis of sediment may be biological, chemical, or physical in nature and may be used to determine the following:

- C toxicity;
- biological availability and effects of contaminants;
- C benthic biota:
- c extent and magnitude of contamination:
- C contaminant migration pathways and source;
- C fate of contaminants:
- C grain size distribution.

The methodologies discussed in this SOP are applicable to the sampling of sediment in both flowing and standing water. They are generic in nature and may be modified in whole or part to meet the handling and analytical requirements of the contaminants of concern, as well as the constraints presented by site conditions and equipment limitations. However, if modifications occur, they should be documented in a site or personal logbook and discussed in reports summarizing field activities and analytical results.

For the purposes of this procedure, sediments are those mineral and organic materials situated beneath an aqueous layer. The aqueous layer may be either static, as in lakes, ponds, and impoundments; or flowing, as in rivers and streams.

Mention of trade names or commercial products does not constitute U.S. EPA endorsement or recommendation for use.

2.0 METHOD SUMMARY

Sediment samples may be collected using a variety of methods and equipment, depending on the depth of the aqueous layer, the portion of the sediment profile required (surface vs. subsurface), the type of sample required (disturbed vs. undisturbed), contaminants present, and sediment type.

Sediment is collected from beneath an aqueous layer either directly, using a hand held device such as a shovel, trowel, or auger; or indirectly, using a remotely activated device such as an Ekman or Ponar dredge. Following collection, sediment is transferred from the sampling device to a sample container of appropriate size and construction for the analyses requested. If composite sampling techniques are employed, multiple grabs are placed into a container constructed of inert material, homogenized, and transferred to sample containers appropriate for the analyses requested. The homogenization procedure should not be used if sample analysis includes volatile organics; in this case, sediment, or multiple grabs of sediment, should be transferred directly from the sample collection device or homogenization container to the sample container.

3.0 SAMPLE PRESERVATION, CONTAINERS, HANDLING AND STORAGE

- Chemical preservation of solids is generally not recommended. Cooling to 4°C is usually the best approach, supplemented by the appropriate holding time for the analyses requested.
- Wide mouth glass containers with Teflon lined caps are utilized for sediment samples.
 The sample volume is a function of the analytical requirements and will be specified in the Work Plan.
- If analysis of sediment from a discrete depth or location is desired, sediment is transferred directly from the sampling device to a labeled sample container(s) of appropriate size and construction for the analyses

requested. Transfer is accomplished with a stainless steel or plastic lab spoon or equivalent.

- If composite sampling techniques or multiple grabs are employed, equal portions of sediment from each location are deposited into a stainless steel, plastic, or other appropriate composition (e.g., Teflon) containers. The sediment is homogenized to obtain a composite thoroughly representative of the area sampled. The composite sediment sample is transferred to a labeled container(s) of appropriate size and construction for the analyses requested. Transfer of sediment is accomplished with a stainless steel or plastic lab spoon or equivalent. Samples for volatile organic analysis must be transferred directly from the sample collection device or pooled from multiple areas in the homogenization container prior to mixing. This is done to minimize loss of contaminant due to volatilization during homogenization.
- devices should he 5. sampling decontaminated, then wrapped in aluminum foil. The sampling device should remain in this wrapping until it is needed. Each sampling device should be used for only one sample. Disposable sampling devices for sediment are generally impractical due to cost and the large number of sediment samples which may be required. Sampling devices should be cleaned in the field using the decontamination procedure described in the Sampling Equipment Decontamination SOP.

4.0 INTERFERENCES AND POTENTIAL PROBLEMS

Substrate particle size and organic matter content are a direct consequence of the flow characteristics of a waterbody. Contaminants are more likely to be concentrated in sediments typified by fine particle size and a high organic matter content. This type of sediment is most likely to be collected from depositional zones. In contrast, coarse sediments with low organic matter content do not typically concentrate pollutants and are generally found in erosional zones. The selection of a sampling location

can, therefore, greatly influence the analytical results and should be justified and specified in the Work Plan.

5.0 EQUIPMENT/APPARATUS

Equipment needed for collection of sediment samples may include:

- C Maps/plot plan
- C Safety equipment
- C Compass
- C Tape measure
- C Survey stakes, flags, or buoys and anchors
- Camera and film
- C Stainless steel, plastic, or other appropriate composition bucket
- 4-oz., 8-oz., and one-quart wide mouth jars w/Teflon lined lids
- C Ziploc plastic bags
- C Logbook
- C Sample jar labels
- Chain of Custody records, field data sheets
- Cooler(s)
- C Ice
- C Decontamination supplies/equipment
- C Spade or shovel
- C Spatula
- C Scoop
- C Trowel
- C Bucket auger
- C Tube auger
- C Extension rods
- C "T" handle
- C Sediment coring device (tube, drive head, eggshell check value, nosecone, acetate tube, extension rods, "T" handle)
- C Ponar dredge
- C Ekman dredge
- C Nylon rope or steel cable
- C Messenger device

6.0 REAGENTS

Reagents are not used for preservation of sediment samples. Decontamination solutions are specified in the Sampling Equipment Decontamination SOP.

7.0 PROCEDURES

7.1 Preparation

- 1. Determine the objective(s) and extent of the sampling effort. The sampling methods to be employed, and the types and amounts of equipment and supplies required will be a function of site characteristics and objectives of the study.
- Obtain the necessary sampling and monitoring equipment.
- Prepare schedules, and coordinate with staff, client, and regulatory agencies, if appropriate.
- 4. Decontaminate or preclean equipment, and ensure that it is in working order.
- 5. Perform a general site survey prior to site entry in accordance with the site specific Health and Safety Plan.
- 6. Use stakes, flagging, or buoys to identify and mark all sampling locations. Specific site factors including flow regime, basin morphometry, sediment characteristics, depth of overlying aqueous layer, contaminant source, and extent and nature of contamination should be considered when selecting sample locations. If required, the proposed locations may be adjusted based on site access, property boundaries, and surface obstructions.

7.2 Sample Collection

Selection of a sampling device is most often contingent upon: (1) the depth of water at the sampling location, and (2) the physical characteristics of the sediment to be sampled. The following procedures may be utilized:

7.2.1 Sampling Surface Sediment with a Trowel or Scoop from Beneath a Shallow Aqueous Layer

For the purpose of this method, surface sediment is considered to range from 0 to six inches in depth and a shallow aqueous layer is considered to range from 0 to 12 inches in depth. Collection of surface sediment from beneath a shallow aqueous layer can be accomplished with tools such as spades, shovels, trowels, and scoops. Although this method can be

used to collect both unconsolidated/consolidated sediment, it is limited somewhat by the depth and movement of the aqueous layer. Deep and rapidly flowing water render this method less accurate than others discussed below. However, representative samples can be collected with this procedure in shallow sluggish water provided care is demonstrated by the sample team member. A stainless steel or plastic sampling implement will suffice in most applications. Care should be exercised to avoid the use of devices plated with chrome or other materials; plating is particularly common with garden trowels.

The following procedure will be used to collect sediment with a scoop, shovel, or trowel:

- Using a decontaminated sampling implement, remove the desired thickness and volume of sediment from the sampling area.
- Transfer the sample into an appropriate sample or homogenization container. Ensure that non-dedicated containers have been adequately decontaminated.
- Surface water should be decanted from the sample or homogenization container prior to sealing or transfer; care should be taken to retain the fine sediment fraction during this procedure.
- 7.2.2 Sampling Surface Sediment with a Bucket Auger or Tube Auger from Beneath a Shallow Aqueous Layer

For the purpose of this method, surface sediment is considered to range from 0 to six inches in depth and a shallow aqueous layer is considered to range from 0 to 24 inches in depth. Collection of surface sediment from beneath a shallow aqueous layer can be accomplished with a system consisting of bucket auger or tube auger, a series of extensions, and a "T" handle (Figure 1, Appendix A). The use of additional extensions in conjunction with a bucket auger can increase the depth of water from which sediment can be collected from 24 inches to 10 feet or more. However, sample handling and manipulation increases in difficulty with increasing depth of water. The bucket auger or tube auger is driven into the sediment and used to extract a core. The various depths represented by the core are homogenized or a subsample of the core is taken from the appropriate depth.

The following procedure will be used to collect sediment samples with a bucket auger or tube auger:

- An acetate core may be inserted into the bucket auger or tube auger prior to sampling if characteristics of the sediments or waterbody warrant. By using this technique, an intact core can be extracted.
- 2. Attach the auger head to the required length of extensions, then attach the "T" handle to the upper extension.
- 3. Clear the area to be sampled of any surface debris.
- 4. Insert the bucket auger or tube auger into the sediment at a 0° to 20° angle from vertical. This orientation minimizes spillage of the sample from the sampler upon extraction from the sediment and water.
- 5. Rotate the auger to cut a core of sediment.
- 6. Slowly withdraw the auger; if using a tube auger, make sure that the slot is facing upward.
- Transfer the sample or a specified aliquot of sample into an appropriate sample or homogenization container. Ensure that nondedicated containers have been adequately decontaminated.
- 7.2.3 Sampling Deep Sediment with a Bucket Auger or Tube Auger from Beneath a Shallow Aqueous Layer

For the purpose of this method, deep sediment is considered to range from six to greater than 18 inches in depth and a shallow aqueous layer is considered to range from 0 to 24 inches. Collection of deep sediment from beneath a shallow aqueous layer can be accomplished with a system consisting of a bucket auger, a tube auger, a series of extensions and a "T" handle. The use of additional extensions can increase the depth of water from which sediment can be collected from 24 inches to five feet or more. However, water clarity must be high enough to permit the sampler to directly observe the sampling

operation. In addition, sample handling and manipulation increases in difficulty with increasing depth of water. The bucket auger is used to bore a hole to the upper range of the desired sampling depth and then withdrawn. The tube auger is then lowered down the borehole, and driven into the sediment to the lower range of the desired sampling depth. The tube is then withdrawn and the sample recovered from the tube. This method can be used to collect firmly consolidated sediments, but is somewhat limited by the depth of the aqueous layer, and the integrity of the initial borehole.

The following procedure will be used to collect deep sediment samples with a bucket auger and a tube auger:

- 1. Attach the bucket auger bit to the required lengths of extensions, then attach the "T" handle to the upper extension.
- Clear the area to be sampled of any surface debris.
- Begin augering, periodically removing any accumulated sediment (i.e., cuttings) from the auger bucket. Cuttings should be disposed of far enough from the sampling area to minimize cross contamination of various depths.
- 4. After reaching the upper range of the desired depth, slowly and carefully remove bucket auger from the boring.
- 5. Attach the tube auger bit to the required lengths of extensions, then attach the "T" handle to the upper extension.
- 6. Carefully lower tube auger down borehole using care to avoid making contact with the borehole sides and, thus, cross contaminating the sample. Gradually force tube auger into sediment to the lower range of the desired sampling depth. Hammering of the tube auger to facilitate coring should be avoided as the vibrations may cause the boring walls to collapse.
- Remove tube auger from the borehole, again taking care to avoid making contact with the borehole sides and, thus, cross contaminating

the sample.

- Discard the top of core (approximately 1 inch); as this represents material collected by the tube auger before penetration to the layer of concern.
- Transfer sample into an appropriate sample or homogenization container. Ensure that non-dedicated containers have been adequately decontaminated.
- 7.2.4 Sampling Surface Sediment with an Ekman or Ponar Dredge from Beneath a Shallow or Deep Aqueous Layer

For the purpose of this method, surface sediment is considered to range from 0 to six inches in depth. Collection of surface sediment can be accomplished with a system consisting of a remotely activated device (dredge) and a deployment system. This technique consists of lowering a sampling device (dredge) to the surface of the sediment by use of a rope, cable, or extended handle. The mechanism is activated, and the device entraps sediment in spring loaded or lever operated jaws.

An Ekman dredge is a lightweight sediment sampling device with spring activated jaws. It is used to collect moderately consolidated, fine textured sediment. The following procedure will be used for collecting sediment with an Ekman dredge (Figure 2, Appendix A):

- 1. Attach a sturdy nylon rope or stainless steel cable through the hole on the top of the bracket, or secure the extension handle to the bracket with machine bolts.
- 2. Attach springs to both sides of the jaws. Fix the jaws so that they are in open position by placing trip cables over the release studs. Ensure that the hinged doors on the dredge top are free to open.
- 3. Lower the sampler to a point 4 to 6 inches above the sediment surface.
- Drop the sampler to the sediment.
- 5. Trigger the jaw release mechanism by

lowering a messenger down the line, or by depressing the button on the upper end of the extension handle.

- 6. Raise the sampler and slowly decant any free liquid through the top of the sampler. Care should be taken to retain the fine sediment fraction during this procedure.
- 7. Open the dredge jaws and transfer the sample into a stainless steel, plastic or other appropriate composition (e.g., Teflon) container. Ensure that non-dedicated containers have been adequately decontaminated. If necessary, continue to collect additional sediment grabs until sufficient material has been secured to fulfill analytical requirements. Thoroughly homogenize and then transfer sediment to sample containers appropriate for the analyses requested. Samples for volatile organic analysis must be collected directly from the bucket before homogenization to minimize volatilization of contaminants.

A Ponar dredge is a heavyweight sediment sampling device with weighted jaws that are lever or spring activated. It is used to collect consolidated fine to coarse textured sediment. The following procedure will be used for collecting sediment with a Ponar dredge (Figure 3, Appendix A):

- 1. Attach a sturdy nylon rope or steel cable to the ring provided on top of the dredge.
- Arrange the Ponar dredge with the jaws in the open position, setting the trip bar so the sampler remains open when lifted from the top. If the dredge is so equipped, place the spring loaded pin into the aligned holes in the trip bar.
- 3. Slowly lower the sampler to a point approximately two inches above the sediment.
- 4. Drop the sampler to the sediment. Slack on the line will release the trip bar or spring loaded pin; pull up sharply on the line closing the dredge.
- 5. Raise the dredge to the surface and slowly

decant any free liquid through the screens on top of the dredge. Care should be taken to retain the fine sediment fraction during this operation.

6. Open the dredge and transfer the sediment to a stainless steel, plastic or other appropriate composition (e.g., Teflon) container. Ensure that non-dedicated containers have been adequately decontaminated. If necessary, continue to collect additional sediment until sufficient material has been secured to fulfill analytical requirements. Thoroughly homogenized and then transfer sediment to sample containers appropriate for the analyses requested. Samples for volatile organic analysis must be collected directly from the bucket before homogenization to minimize volatilization of contaminants.

7.2.5 Sampling Subsurface Sediment with a Coring Device from Beneath a Shallow Aqueous Layer

For purposes of this method, subsurface sediment is considered to range from 6 to 24 inches in depth and a shallow aqueous layer is considered to range from 0 to 24 inches in depth. Collection of subsurface sediment from beneath a shallow aqueous layer can be accomplished with a system consisting of a tube sampler, acetate tube, eggshell check valve, nosecone, extensions, and "T" handle, or drivehead. The use of additional extensions can increase the depth of water from which sediment can be collected from 24 inches to 10 feet or more. This sampler may be used with either a drive hammer for firm sediment, or a "T" handle for soft sediment. However, sample handling and manipulation increases in difficulty with increasing depth of water.

The following procedure describes the use of a sample coring device (Figure 4, Appendix A) used to collect subsurface sediments.

- 1. Assemble the coring device by inserting the acetate core into the sampling tube.
- 2. Insert the "egg shell" check valve into the lower end of the sampling tube with the convex surface positioned inside the acetate core.

- Screw the nosecone onto the lower end of the sampling tube, securing the acetate tube and eggshell check valve.
- 4. Screw the handle onto the upper end of the sampling tube and add extension rods as needed.
- 5. Place the sampler in a perpendicular position on the sediment to be sampled.
- 6. If the "T" handle is used, place downward pressure on the device until the desired depth is reached. After the desired depth is reached, rotate the sampler to shear off the core at the bottom. Slowly withdraw the sampler from the sediment and proceed to Step 15.
- 7. If the drive hammer is selected, insert the tapered handle (drive head) of the drive hammer through the drive head.
- 8. Drive the sampler into the sediment to the desired depth.
- 9. Record the length of the tube that penetrated the sample material, and the number of blows required to obtain this depth.
- 10. Remove the drive hammer and fit the keyhole-like opening on the flat side of the hammer onto the drive head. In this position, the hammer serves as a handle for the sampler.
- 11. Rotate the sampler to shear off the core at the bottom.
- 12. Lower the sampler handle (hammer) until it just clears the two ear-like protrusions on the drive head, and rotate about 90°.
- 13. Slowly withdraw the sampler from the sediment. If the drivehead was used, pull the hammer upwards and dislodge the sampler from the sediment.
- 14. Carefully remove the coring device from the water.
- 15. Unscrew the nosecone and remove the

eggshell check valve.

- 16. Slide the acetate core out of the sampler tube. Decant surface water, using care to retain the fine sediment fraction. If head space is present in the upper end, a hacksaw may be used to shear the acetate tube off at the sediment surface. The acetate core may then be capped at both ends. Indicate on the acetate tube the appropriate orientation of the sediment core using a waterproof marker. The sample may be used in this fashion, or the contents transferred to a sample or homogenization container.
- 17. Open the acetate tube and transfer the sediment to a stainless steel, plastic or other appropriate composition (e.g., Teflon) container. Ensure that non-dedicated containers have heen adequately decontaminated. If necessary, continue to collect additional sediment until sufficient material has been secured to fulfill analytical requirements. Thoroughly homogenize and then transfer sediment to sample containers appropriate for the analyses requested. Samples for volatile organic analysis must be collected directly from the bucket before homogenization to minimize volatilization of contaminants.

More specifically, when sampling sediment from waterbodies, physical hazards must be identified and adequate precautions must be taken to ensure the safety of the sampling team. The team member collecting the sample should not get too close to the edge of the waterbody, where bank failure may cause loss of balance. To prevent this, the person performing the sampling should be on a lifeline, and be wearing adequate protective equipment. If sampling from a vessel is determined to be necessary, appropriate protective measures must be implemented.

12.0 REFERENCES

Mason, B.J., Preparation of Soil Sampling Protocol:

8.0 CALCULATIONS

This section is not applicable to this SOP.

9.0 QUALITY ASSURANCE/ QUALITY CONTROL

There are no specific quality assurance (QA) activities which apply to the implementation of these procedures. However, the following QA procedures apply:

- 1. All data must be documented on field data sheets or within site logbooks.
- 2. All instrumentation must be operated in accordance with operating instructions as supplied by the manufacturer, unless otherwise specified in the work plan. Equipment checkout and calibration activities must occur prior to sampling/operation, and they must be documented.

10.0 DATA VALIDATION

This section is not applicable to this SOP.

11.0 HEALTH AND SAFETY

When working with potentially hazardous materials, follow U.S. EPA/OSHA and Corporate health and safety procedures.

Technique and Strategies. 1983 EPA-600/4-83-020.

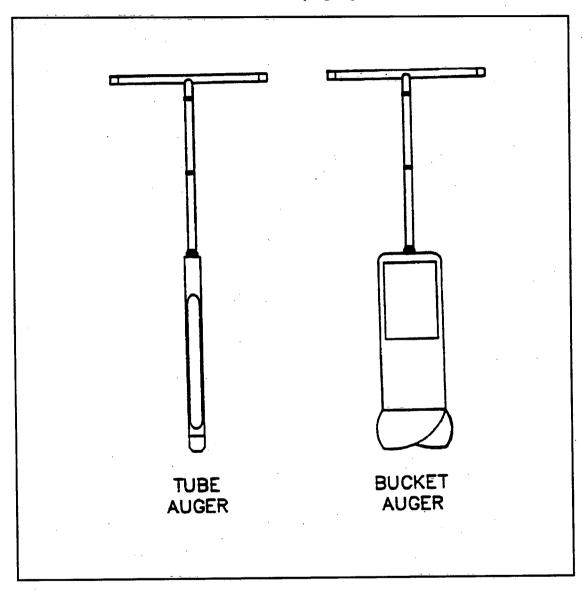
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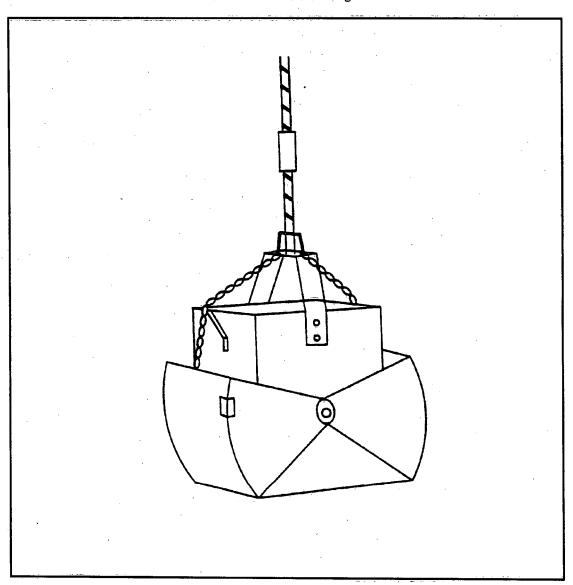
APPENDIX A

FIGURE 1. Sampling Auger



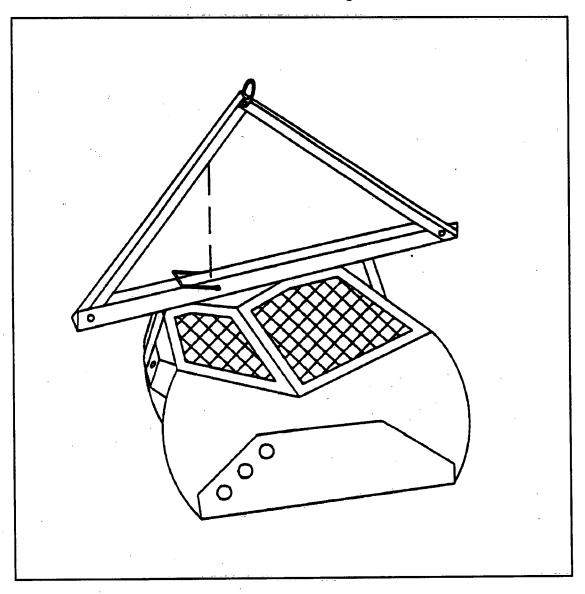
APPENDIX A (Cont'd)

FIGURE 2. Ekman Dredge



APPENDIX A (Cont'd)

FIGURE 3. Ponar Dredge



APPENDIX A (Cont'd)

FIGURE 4. Sample Coring Device

